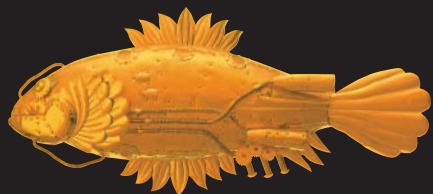


# Shodex™



## HPLC Columns

### Food analysis with Shodex™ columns

*Saccharides, Organic acids, Vitamins, Fatty acids and Amino acids*

**Technical notebook**

**No. 3**



**SHOWA  
DENKO**  
EUROPE

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## 1. Introduction

In recent years, a broad range of analyses, including agrochemical residues and food additives, have been performed on foods with increasingly greater concern about their safety. This article describes analyses of general food ingredients, especially those based on high performance liquid chromatography (HPLC).

## 2. Saccharides

HPLC-based analyses of saccharides and sugar alcohols are performed in various modes, such as normal phase, ligand exchange, size exclusion, and ion exchange. There are two series of Shodex columns for saccharide analysis: those operated mainly in ligand exchange mode (SUGAR series), and those in normal phase mode (Asahipak NH2P-50 polymeric amino columns). In the SUGAR series columns, a hard styrene-divinylbenzene copolymer designed for saccharide analysis is used as the base material, and a strong cation exchange resin incorporating functional sulfo-groups coupled with metal counter ions is used as the packing. Columns of the SUGAR series are available in 14 types according to counter ion, pore size, and pore structure, showing differential selectivity for different saccharides. On the other hand, the Asahipak NH2P-50 columns comprise a hydrophilic polymer gel [poly(vinyl)alcohol] and a polyamine that is chemically and stably bound to the gel. As such, these columns have excellent durability without the problem of time-related deterioration in silica-based amino columns. Both these series are suitable for saccharide separation and analysis of foods, areas of biochemistry, and natural substances. Other items include the Asahipak GS-220 HQ polymeric packed columns for size exclusion mode to analyze water-soluble, low molecular weight dietary fibers, and the OHPak SB-800 HQ polymeric packed columns for size exclusion mode to analyze polysaccharides, as well as the SUGAR KS-800 columns. Table 1 shows the lineup of Shodex packed columns for saccharide analysis.

**Table 2-1. Columns for Saccharide Analysis (1)**

Product Code	Product Name	Counter Ion	Separation Mode*1	ExclusionLimit (Pullulan)	Theoretical Plate Number (TP/column)	Particle Size (µm)	ID x Length (mm)
F6378100	SUGAR SH1011	H <sup>+</sup>	SEC + IEX	1,000	≥17,000	6	8.0 x 300
F6378101	SUGAR SH1821	H <sup>+</sup>	SEC + IEX	10,000	≥17,000	6	8.0 x 300
F6700080	SUGAR SH-G	H <sup>+</sup>	-	Guard column	-	10	6.0 x 50
F6378102	SUGAR SC1011	Ca <sup>2+</sup>	SEC + LEX	1,000	≥13,000	6	8.0 x 300
F6378103	SUGAR SC1821	Ca <sup>2+</sup>	SEC + LEX	10,000	≥13,000	6	8.0 x 300
F6700090	SUGAR SC-LG	Ca <sup>2+</sup>	-	Guard column	-	10	6.0 x 50
F6378105	SUGAR SP0810	Pb <sup>2+</sup>	SEC + LEX	1,000	≥11,000	7	8.0 x 300
F6700081	SUGAR SP-G	Pb <sup>2+</sup>	-	Guard column	-	10	6.0 x 50
F7001400	SUGAR SC1211	Ca <sup>2+</sup>	NP + LEX	-	≥ 5,500	6	6.0 x 250
F6700120	SUGAR SC-G	Ca <sup>2+</sup>	-	Guard column	-	10	4.6 x 10
F7001300	SUGAR SZ5532	Zn <sup>2+</sup>	NP + LEX	-	≥ 5,500	6	6.0 x 150
F6700110	SUGAR SZ-G	Zn <sup>2+</sup>	-	Guard column	-	6	4.6 x 10
F6378010	SUGAR KS-801	Na <sup>+</sup>	SEC + LEX	1,000	≥17,000	6	8.0 x 300
F6378020	SUGAR KS-802	Na <sup>+</sup>	SEC + LEX	10,000	≥17,000	6	8.0 x 300
F6378025	SUGAR KS-803	Na <sup>+</sup>	SEC	50,000	≥17,000	6	8.0 x 300
F6378035	SUGAR KS-804	Na <sup>+</sup>	SEC	400,000	≥17,000	7	8.0 x 300
F6378050	SUGAR KS-805	Na <sup>+</sup>	SEC	5,000,000	≥ 9,000	17	8.0 x 300
F6378060	SUGAR KS-806	Na <sup>+</sup>	SEC	(50,000,000)*2	≥ 9,000	17	8.0 x 300
F6700020	SUGAR KS-G	Na <sup>+</sup>	-	Guard column	-	10	6.0 x 50
F6378070	SUGAR KS-807	Na <sup>+</sup>	SEC	(200,000,000)*2	≥ 4,000	30	8.0 x 300
F6700021	SUGAR KS-807G	Na <sup>+</sup>	-	Guard column	-	30	8.0 x 50
F7600005	Asahipak GS-220 HQ	-	SEC	3,000	≥19,000	6	7.5 x 300
F6710019	Asahipak GS-2G 7B	-	-	Guard column	-	9	7.5 x 50
F7630002	Asahipak NH2P-50 4D	-	NP	-	≥ 5,000	5	4.6 x 150
F7630001	Asahipak NH2P-50 4E	-	NP	-	≥ 7,500	5	4.6 x 250
F6710016	Asahipak NH2P-50G 4A	-	-	Guard column	-	5	4.6 x 10
F7630006	Asahipak NH2P-50 2D	-	NP	-	≥ 3,500	5	2.0 x 150
F6713000	Asahipak NH2P-50G 2A	-	-	Guard column	-	5	2.0 x 10
F6710100	Asahipak NH2P-LF	-	-	Line filter	-	-	8.0 x 75
F6379230	USPpak MN-431	Ca <sup>2+</sup>	SEC + LEX	-	≥ 4,000	-	4.0 x 250

\*1 : SEC (Size exclusion), IEX (Ion exclusion), LEX (Ligand exchange), NP (Normal phase)

\*2 : Figures in ( ) are estimated values

Table 2-2. Columns for Saccharide Analysis (2)

Product Code	Product Name	Exclusion Limit (Pullulan)	Usable Organic Solvents (Max.%)			Theoretical PlateNumber (TP/column)	Particle Size (μm)	ID x Length (mm)
			Methanol	Acetonitrile	DMF			
F6429100	OHpak SB-802 HQ	4,000	0	0	0	≥12,000	8	8.0 x 300
F6429101	OHpak SB-802.5 HQ	10,000	100	75	100	≥16,000	6	8.0 x 300
F6429102	OHpak SB-803 HQ	100,000	100	75	100	≥16,000	6	8.0 x 300
F6429103	OHpak SB-804 HQ	1,000,000	75	75	100	≥16,000	10	8.0 x 300
F6429104	OHpak SB-805 HQ	4,000,000	75	75	100	≥12,000	13	8.0 x 300
F6429105	OHpak SB-806 HQ	(20,000,000)* <sup>1</sup>	75	75	100	≥12,000	13	8.0 x 300
F6429106	OHpak SB-806M HQ	(20,000,000)* <sup>1</sup>	75	75	100	≥12,000	13	8.0 x 300
F6709430	OHpak SB-G	Guard column	75	75	100	-	10	6.0 x 50
F6429108	OHpak SB-807 HQ	(500,000,000)* <sup>1</sup>	75	75	100	≥15,000	35	8.0 x 300
F6709431	OHpak SB-807 G	Guard column	75	75	100	-	35	6.0 x 50

\*1 : Figures in ( ) are estimated values

## 2-1. Separation Mechanism

### 2-1-1. Anomer Separation of Saccharides

Reducing saccharides can adopt various cyclic structures, as well as a linear structure. In such cases, two tautomers, α type and β type, are produced because carbonyl carbon atoms become asymmetric. The relation between α type and β type is called anomer. (Fig. 2-1)

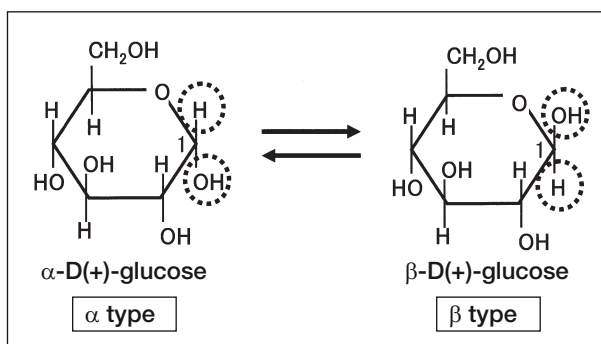


Fig. 2-1 Anomers

Under conditions in which the conversion rate between the tautomers is low, α and β anomers are separated by the column causing the peak tops to split or widen. (Fig. 2-2)

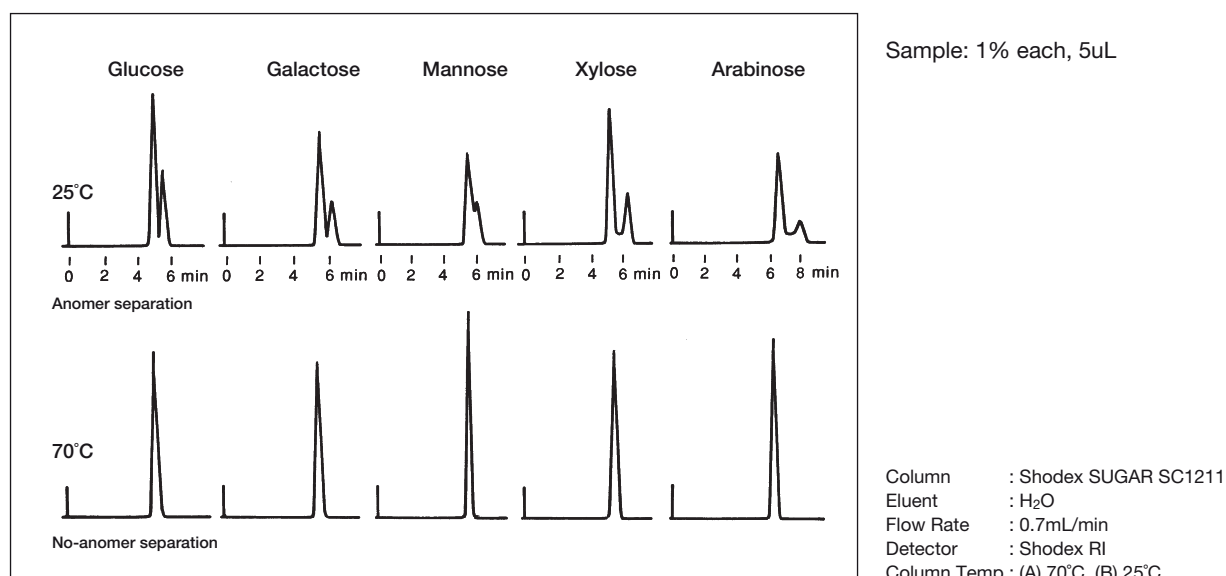


Fig. 2-2 Influence of Anomer Separation in Saccharide Analysis

Two methods are available to prevent anomer separation during analysis:

- \* Analysis at high temperature
- \* Analysis under strong alkaline conditions

### 2-1-2. Shodex SUGAR Series

A strong feature of the Shodex SUGAR series columns resides in the mechanism of separation in the ligand exchange mode. Ligand exchange refers to a mode of separation based on the interaction (ligand exchange potential) between hydroxyl groups and metal ions to form a complex. Saccharides have a 5-membered (furanose) or 6-membered (pyranose) ring structure, containing a large number of hydroxyl groups. These hydroxyl groups bind together either equatorially or axially with respect to the carbon plane. This conformation of the hydroxyl groups differs depending on the kind of saccharide.

Figure 2-3 shows the relationship between hydroxyl group conformation and counter ion interaction.

In Figure 2-3(a), three hydroxyl groups form a complex with the metal ion. In Figure 2-3(b), only two hydroxyl groups form a complex with the metal ion due to the hydroxyl group conformation. Hence, ligand exchange potential is higher for (a) than for (b).

The complex formation potential also differs depending on the kind of metal ion.

\* When using a column of the SUGAR series, analysis should be performed at increased column temperatures to prevent anomer separation. Under strongly alkaline conditions, saccharides are likely to isomerize with the fear of decomposition of polysaccharides.

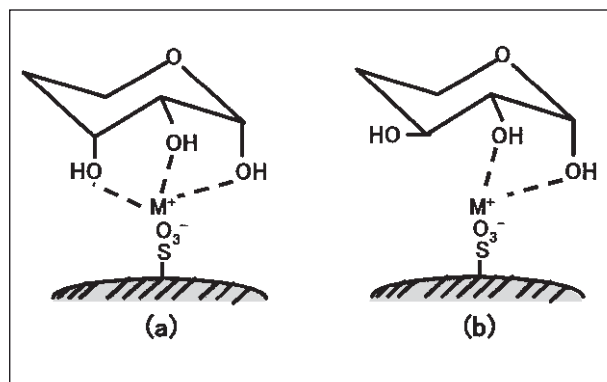


Fig. 2-3 Difference of counter ion interaction

### 2-1-3. Shodex Asahipak NH2P-50 Series

Columns of the Shodex Asahipak NH2P-50 series are greatly improved amino columns which not only maintain the high separation performance of the conventional silica-based amino columns, but also solve the problem of declines in retention over time. This is due to stable chemical bonding of polyamine with hydrophilic polymer gel. Other advantages are:

- \* Analysis under moderate conditions (around pH 7 and room temperature) is possible.
- \* Sharp, near-symmetric peaks can be obtained for a wide variety of saccharides.
- \* Accurate quantitative determination can be made.
- \* A wide range of eluents, such as various buffer solutions, alkaline solutions, or acidic solutions can be used.
- \* Alkaline washing of columns is possible.

With amino columns, saccharides elute in order of increasing polarity due to the function of normal phase chromatography. Usually, a mixed solvent of acetonitrile and water is used as the eluent. When the mixing ratio of acetonitrile is increased, the polarity of the eluent becomes lower. This results in a stronger interaction between saccharides and the column and a larger elution volume.

As NH2P-50 columns have weak alkaline amino groups, the condition inside the column is alkaline. This enables saccharides to be analyzed without causing separation of anomers even at room temperature.

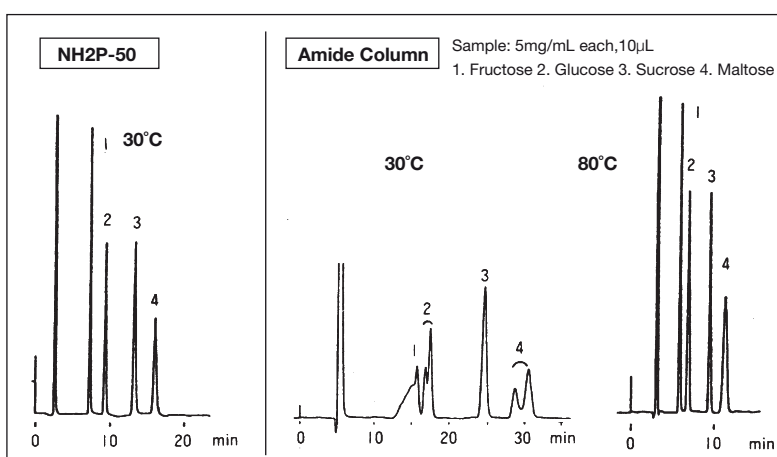


Fig. 2-4 Effects of Column Temperature on Elution Patterns (Comparison with Amide Column)

There are columns, called amide columns, which are used for analysis of saccharides under the same elution conditions as those for amino columns. Although amide columns have acrylamide groups introduced, analysis has to be made at high temperatures because the acrylamide group is not alkaline. (Fig. 2-4)

Column : Shodex Asahipak NH2P-50 4E  
(4.6x250mm)  
Amide Column from Company-A  
(4.6x250mm)  
Eluent : CH<sub>3</sub>CN/H<sub>2</sub>O=75/25  
Flow Rate : 1.0mL/min  
Detector : Shodex RI  
Column Temp.: 30°C, 80°C

NH2P-50 columns show good reproducibility for a long time, because the chemical structure of the packing material is stable. With silica-based amino columns, retention of saccharide weakens over time, resulting in the significant widening of every peak. This clearly indicates deterioration of the column. (Fig. 2-5)

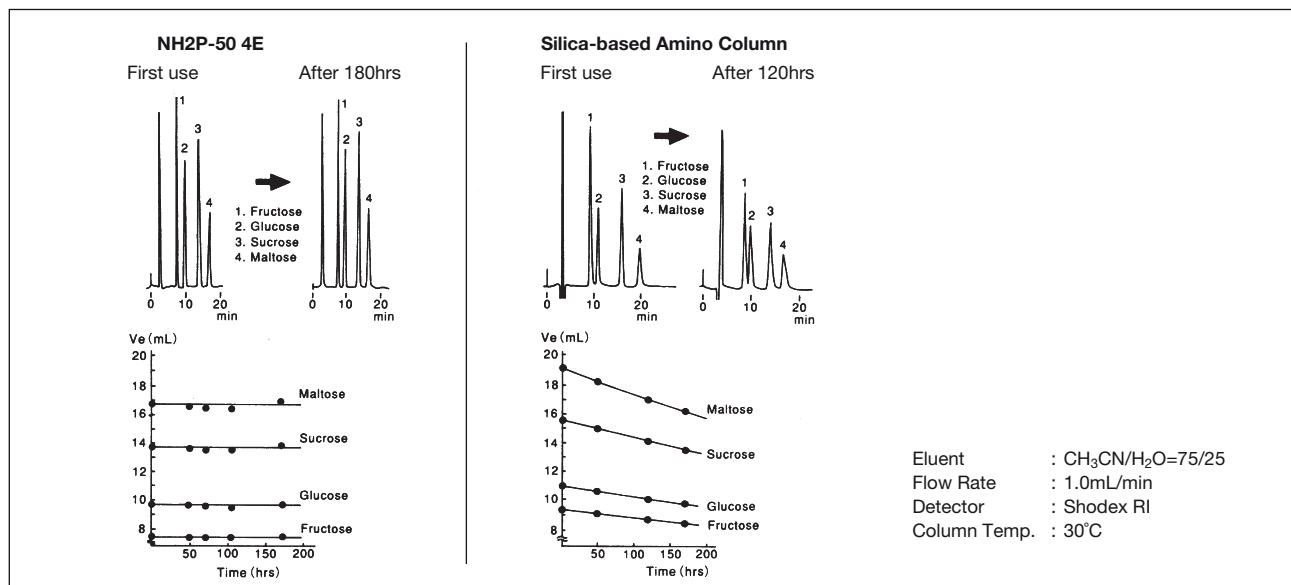


Fig. 2-5 Reproducibility of Chromatograms with NH2P-50 4E Column and Silica-based Amino Column

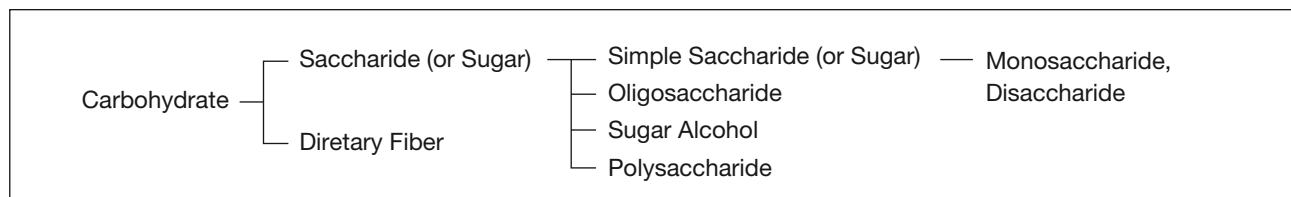
#### 2-1-4. Detection of Saccharides

The most commonly used approach for detecting saccharides is the use of the differential refractive index (RI) detector, which is based on refractive index differences between sample components and a mobile phase. Although the RI detector is highly versatile, it exhibits low selectivity and is not suitable for gradient elution. Another drawback is the tendency for lower sensitivity than other types of detectors. In cases where high selectivity and high sensitivity are required, the ultraviolet absorption (UV) detector and the fluorescence (FL) detector are effective. It should be noted, however, that saccharides must be derivatized because they lack a structure for absorbing UV or emitting fluorescence. Other available methods of detection include the mass spectrum (MS) analyzer, the electrochemical (EC) detector which electrochemically detects ionized saccharides under alkaline conditions, and the evaporation light scattering (ELS) detector which detects saccharides by evaporating an eluent and an irradiating laser.

#### 2-2. Monosaccharides and Disaccharides

The term "Saccharide" generically refers to simple saccharides, oligosaccharides, polysaccharides, sugar alcohols and the like. Simple saccharides, i.e., monosaccharides and disaccharides, exclude sugar alcohols. Emphasized labeling such as, "Very Low Sugar" or "Low Sugar," and "Sugarless" is not permitted unless the content of the relevant type of sugar is not more than 5 grams per 100 grams (2.5 g/100 mL for refreshing beverages), and less than 0.5 grams per 100 grams (0.5 g/100 mL for refreshing beverages), respectively. Japan's nutritional facts labeling standards specify the use of an amino column (4.6 mm inner diameter x 250 mm length) to analyze simple saccharides and oligosaccharides (Figure 2-6).

Shodex can provide many kinds of columns for saccharide analysis.



Monosaccharide, Disaccharide	Oligosaccharide
<p>Column : A column packed with a silica or polymer gel bounded amino (propyl) group (4.6mmID x 250mm) → Shodex Asahipak NH2P-50 4E</p> <p>Eluent : CH<sub>3</sub>CN/H<sub>2</sub>O = 75/25</p> <p>Flow Rate : 1.0 mL/min</p> <p>Detector : RI</p> <p>Column Temp.: Room Temp.</p> <p>Injection Vol. : 20μL</p>	<p>Column : A column packed with a silica or polymer gel bounded amino (propyl) group (4.6mmID x 250mm) → Shodex Asahipak NH2P-50 4E</p> <p>Eluent : CH<sub>3</sub>CN/H<sub>2</sub>O = 75/25</p> <p>Flow Rate : 1.0 mL/min</p> <p>Detector : RI</p> <p>Column Temp.: Room Temp.</p> <p>Injection Vol. : 20μL</p>
Japan's Nutritional Facts Labeling Standards Ver. 3 (Japan Health Food Authorization)	

Fig. 2-6 Analysis Condition of Saccharides and Oligosaccharides

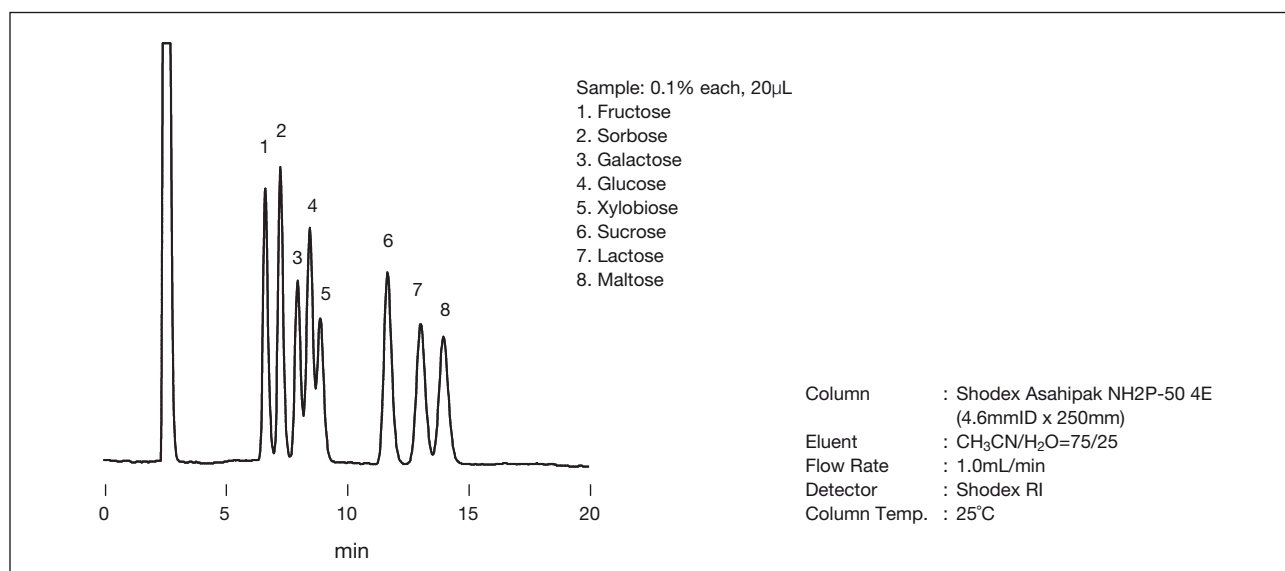


Fig. 2-7 Standard Saccharides with NH2P-50 4E

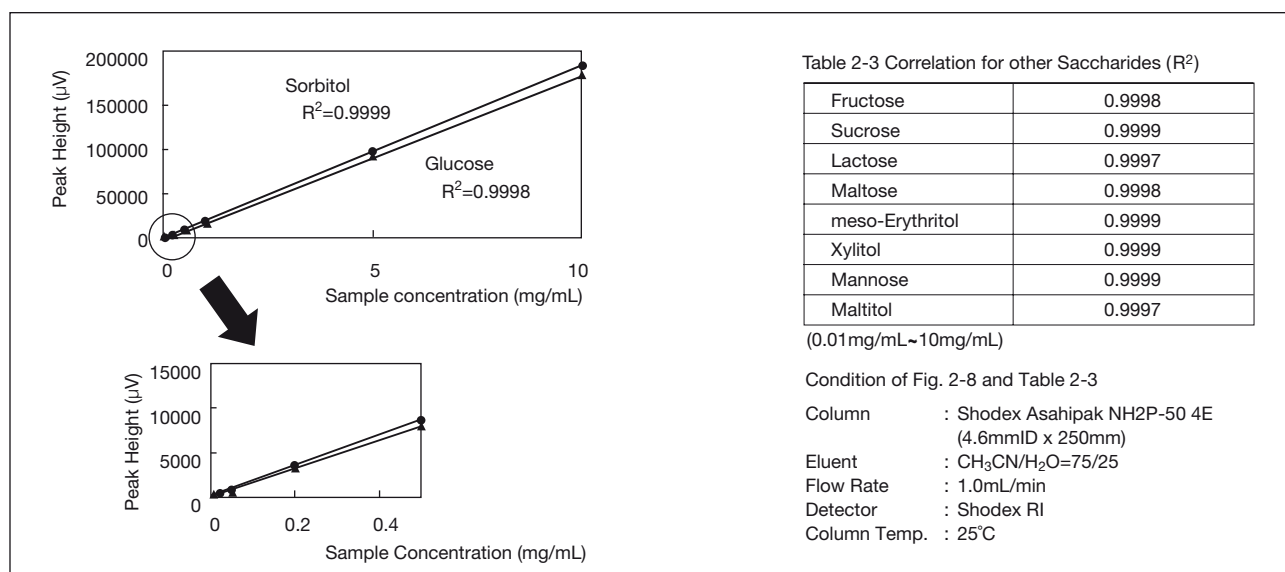


Fig. 2-8 Calibration Curves with NH2P-50



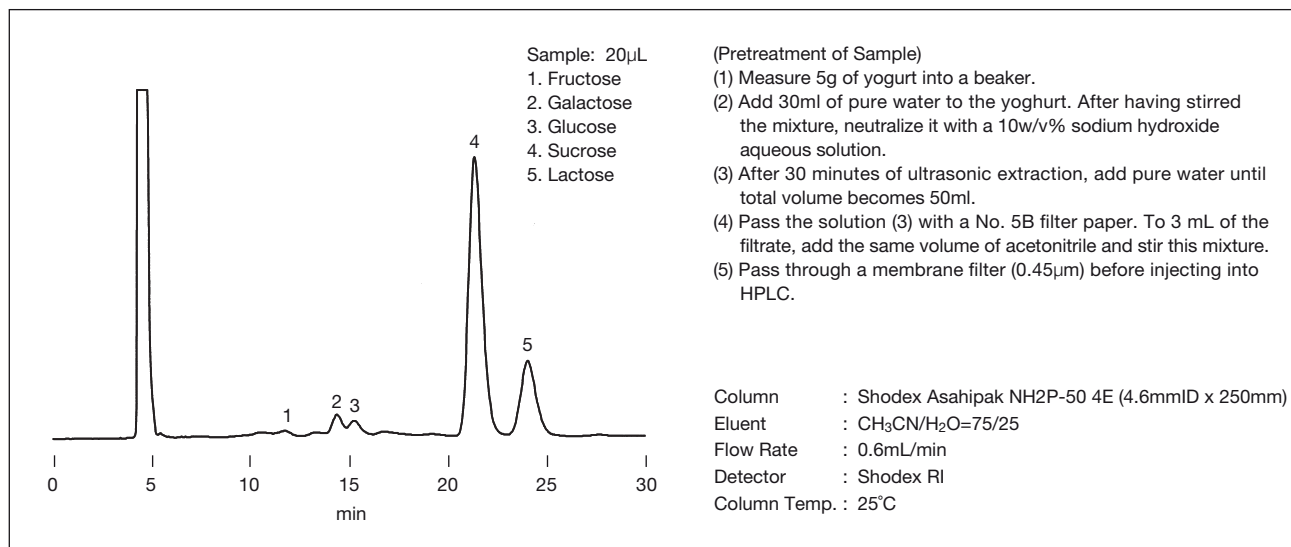


Fig. 2-9 Saccharides in Sugar-added Yoghurt

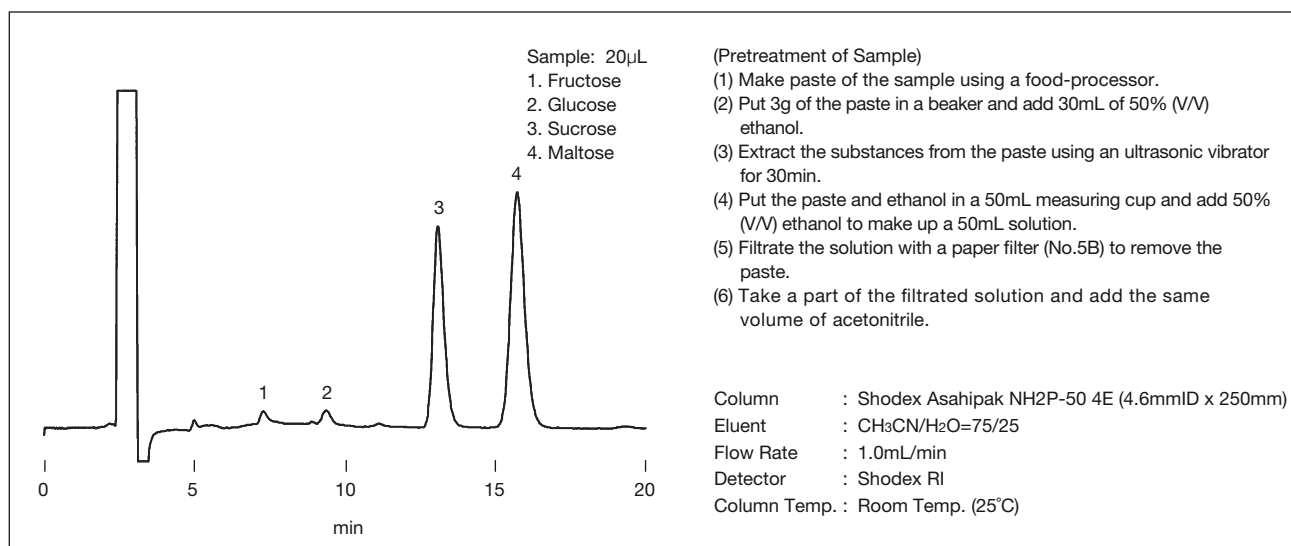


Fig. 2-10 Saccharides in Sweet Potato, Roasted

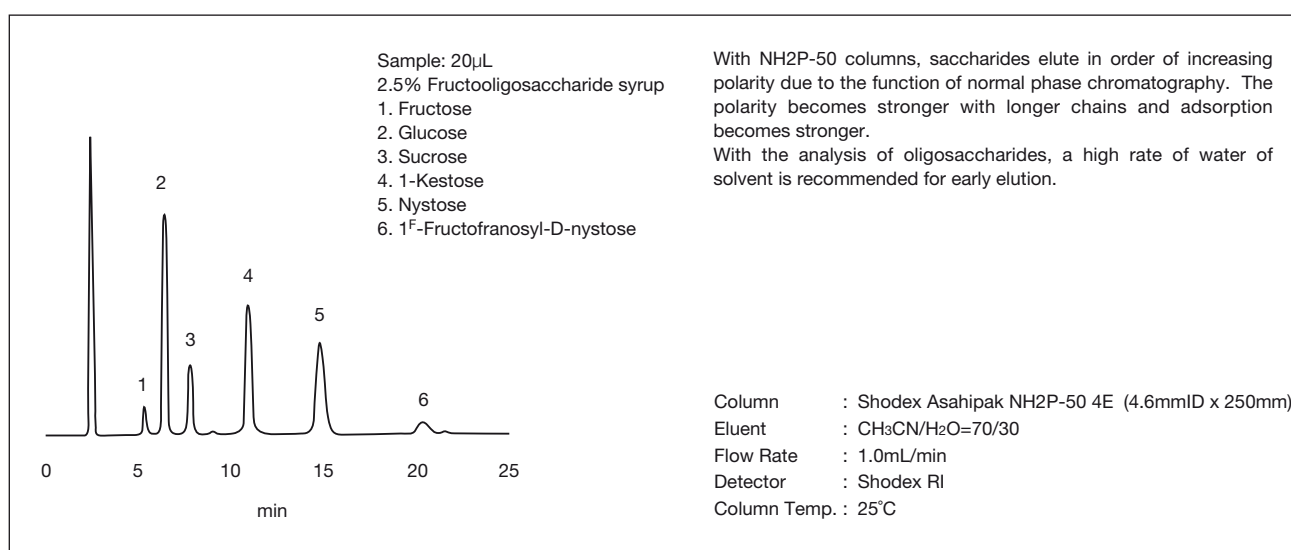


Fig. 2-11 Fructo-oligosaccharide Syrup



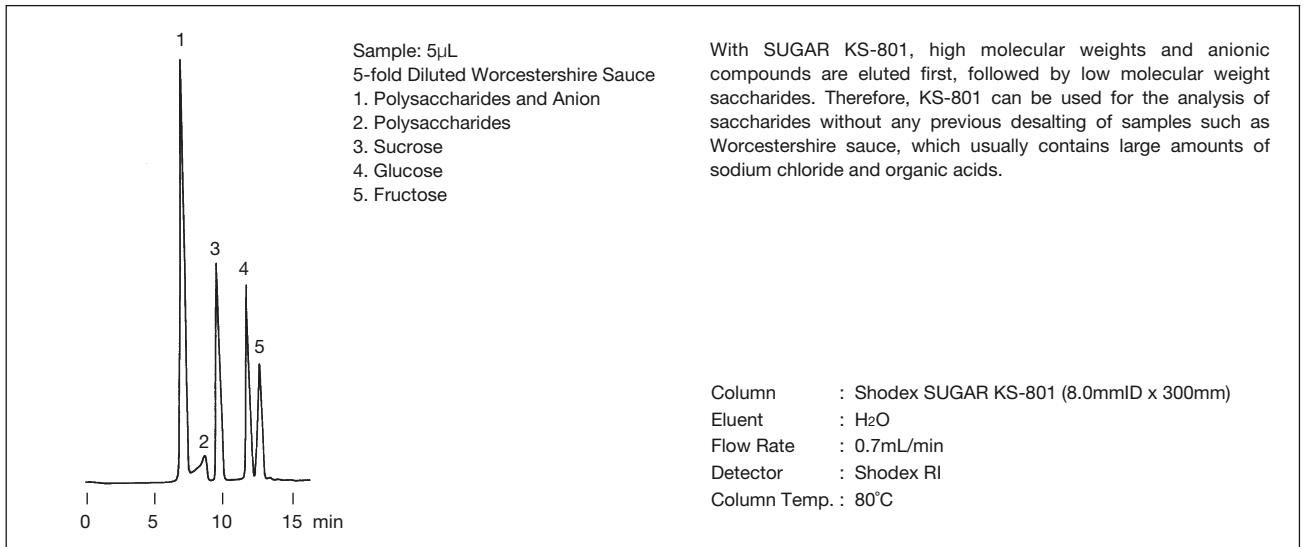


Fig. 2-12 Saccharides in Worcestershire Sauce

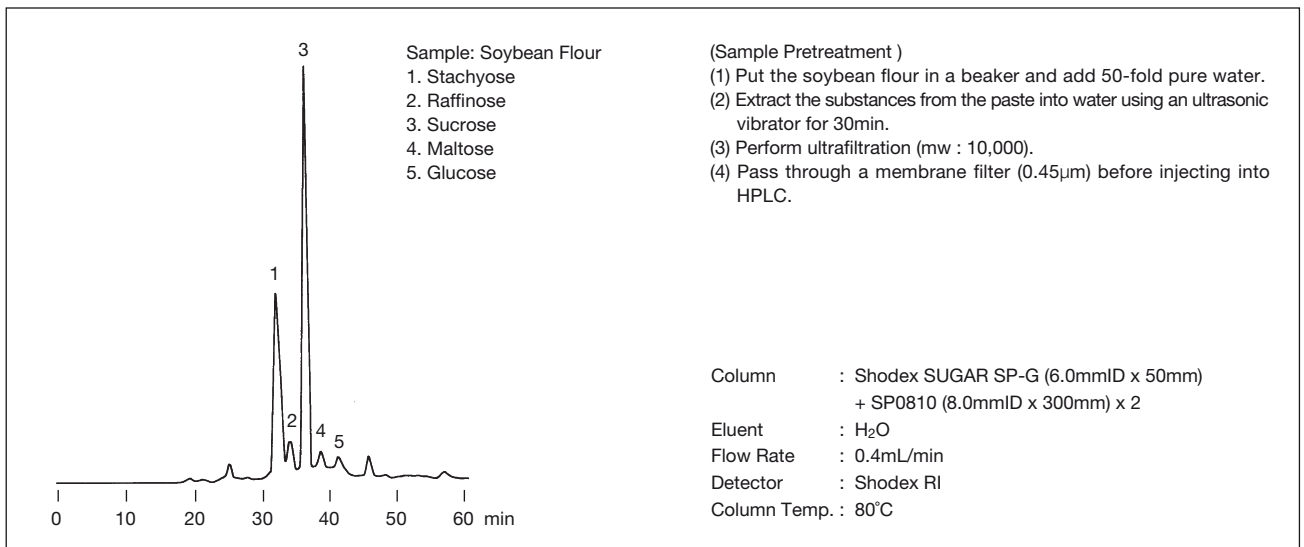


Fig. 2-13 Saccharides in Soybean Flour

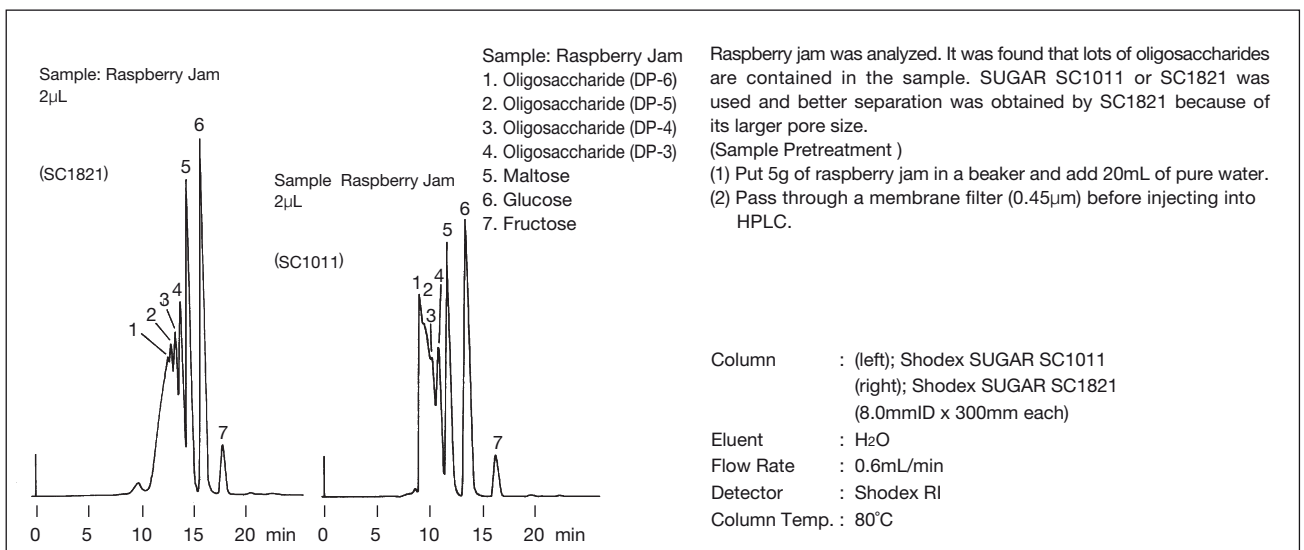


Fig. 2-14 Saccharides in Raspberry Jam

## 2-3. Saccharides and Sugar Alcohols

Amino columns (4.6mmID x 250mm) and ligand exchange mode columns are one of the official methods of analysis for sugar alcohols. (Fig. 2-15)

Sugar Alcohol (1)	Sugar Alcohol (2)
<p>Column : A column packed with a silica or polymer gel bounded amino (propyl) group (4.6mmID x 250mm) → Shodex Asahipak NH2P-50 4E</p> <p>Eluent : CH<sub>3</sub>CN/H<sub>2</sub>O = 75/25</p> <p>Flow Rate : 1.0mL/min</p> <p>Detector : RI</p> <p>Column Temp.: Room Temp.</p> <p>Injection Vol. : 20μL</p>	<p>Column : A column packed with sulfonated poly styrene gel (Pb or Ca ligand) (7.8-8.0mmID x 300mm) → Shodex SUGAR SP0810, SC1011</p> <p>Eluent : H<sub>2</sub>O</p> <p>Flow Rate : 0.6mL/min</p> <p>Detector : RI</p> <p>Column Temp.: 85°C</p> <p>Injection Vol. : 5μL</p>
<p>Japan's Nutritional Facts Labeling Standards Ver. 3 (Japan Health Food Authorization)</p>	

Fig. 2-15 Analysis Condition of Sugar Alcohols

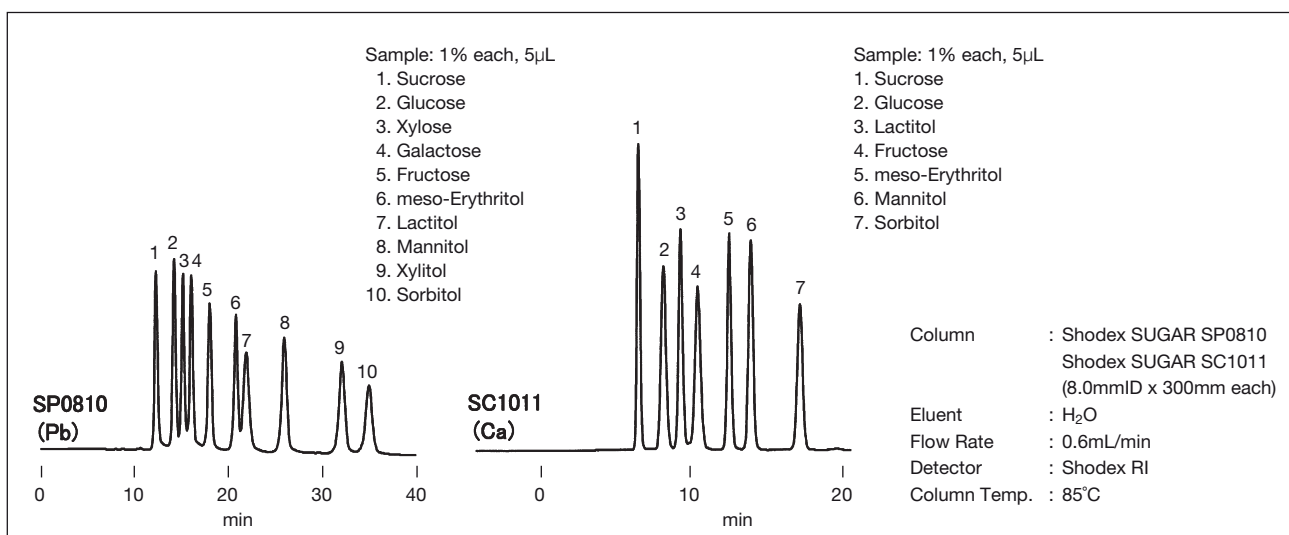


Fig. 2-16 Standard Saccharides and Sugar Alcohol with SP0810, SC1011

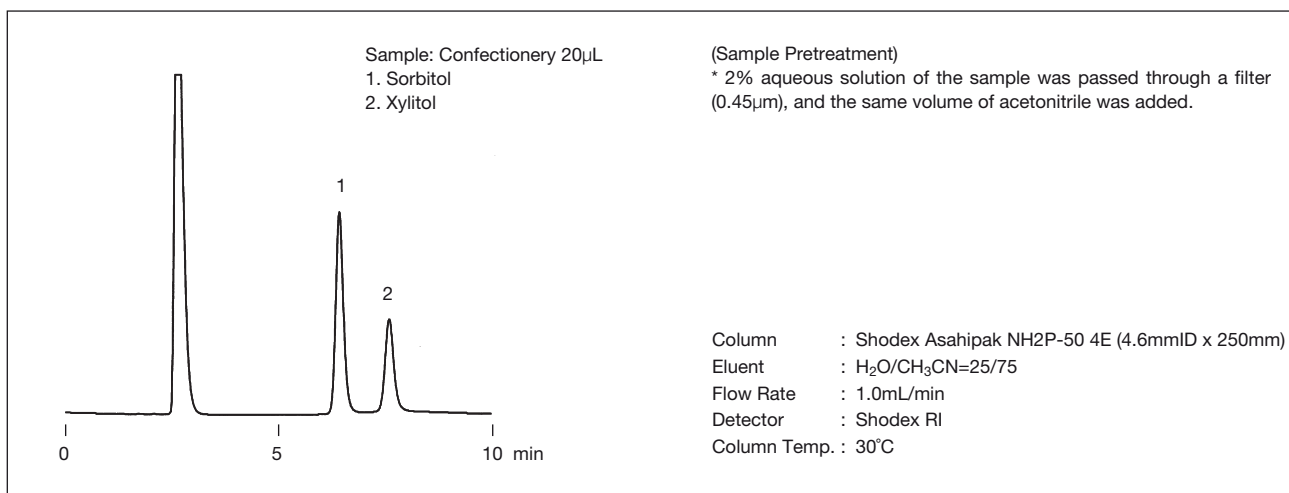


Fig. 2-17 Xylitol in Confectionery

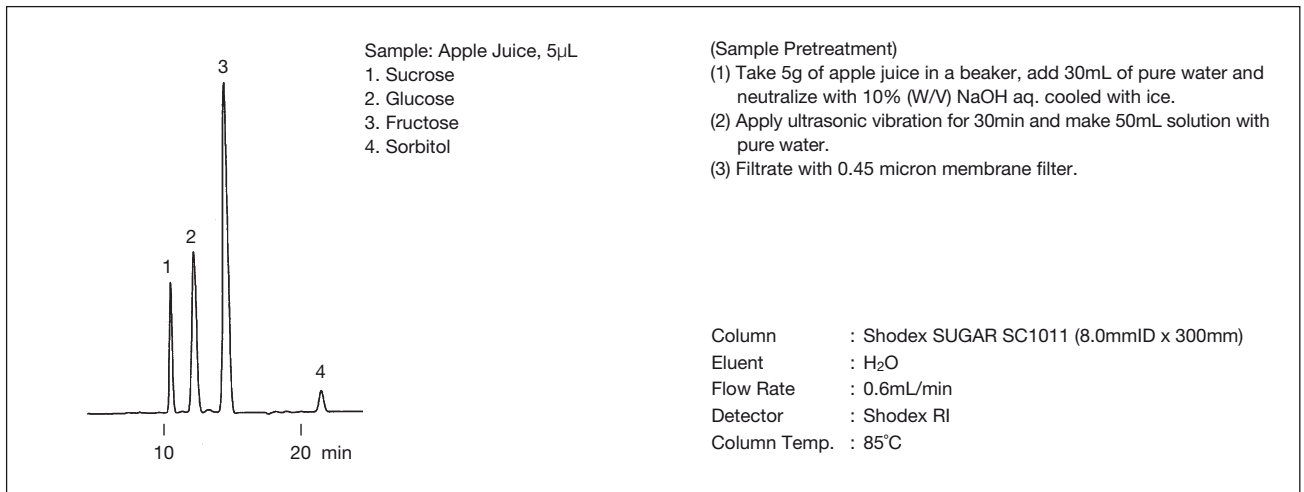


Fig. 2-18 Apple Juice

## 2-4. Water-soluble Low Molecular Weight Dietary Fibers

Dietary fibers are usually quantified by the Prosky method (enzyme-gravimetric method). However, for water-soluble low molecular weight dietary fibers unquantifiable by the Prosky method, the enzyme-HPLC method is used. In this analysis, the sample is divided into two fractions: dietary fibers (tri- and higher saccharides), and mono- and disaccharides. Subsequently, the peak area ratio of the dietary fiber fraction and glucose (or internal standard) is determined.

### Water-soluble Low Molecular Weight Dietary Fibers

Column : Size exclusion chromatography columns  
 (Exclusion limit MW : ca. 5,000)  
 or Ligand exchange columns  
 (Na or Ca type)  
 Eluent : H<sub>2</sub>O  
 Flow Rate : 0.5mL/min  
 Detector : RI  
 Column Temp. : 80°C  
 Injection Vol. : 20 $\mu$ L

Japan's Nutritional Facts Labeling Standards Ver. 3  
 (Japanese Health and Food Nutrition Society)

Fig. 2-19 Analysis Condition of Water Soluble Dietary Fiber

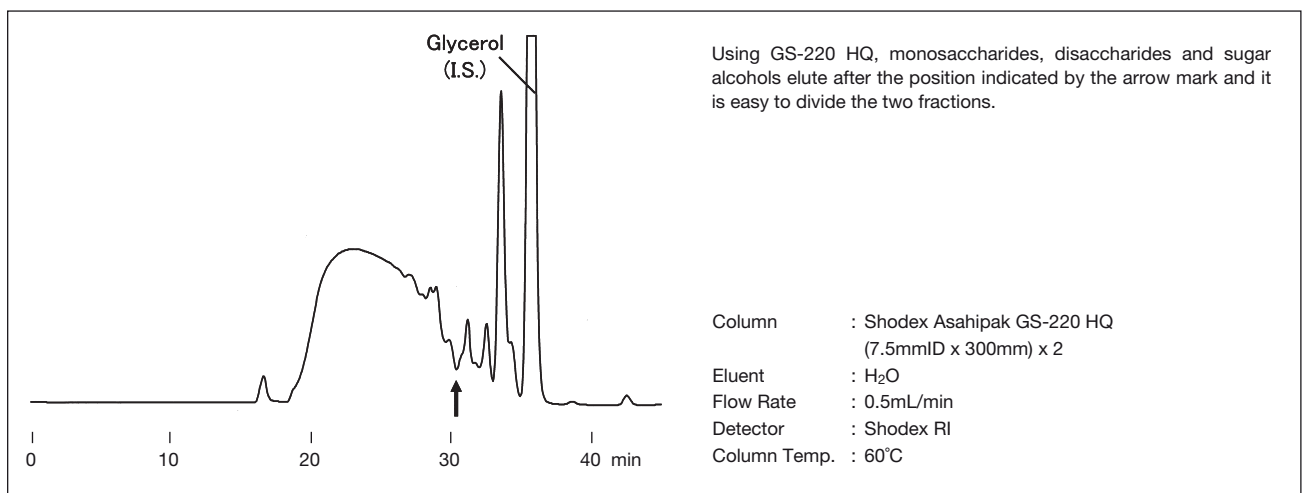


Fig. 2-20 Water-soluble Dietary Fiber

## 2-5. Saccharides with Amino Acids or Organic Acids

Amino acids and organic acids are often contained with saccharides in foods.

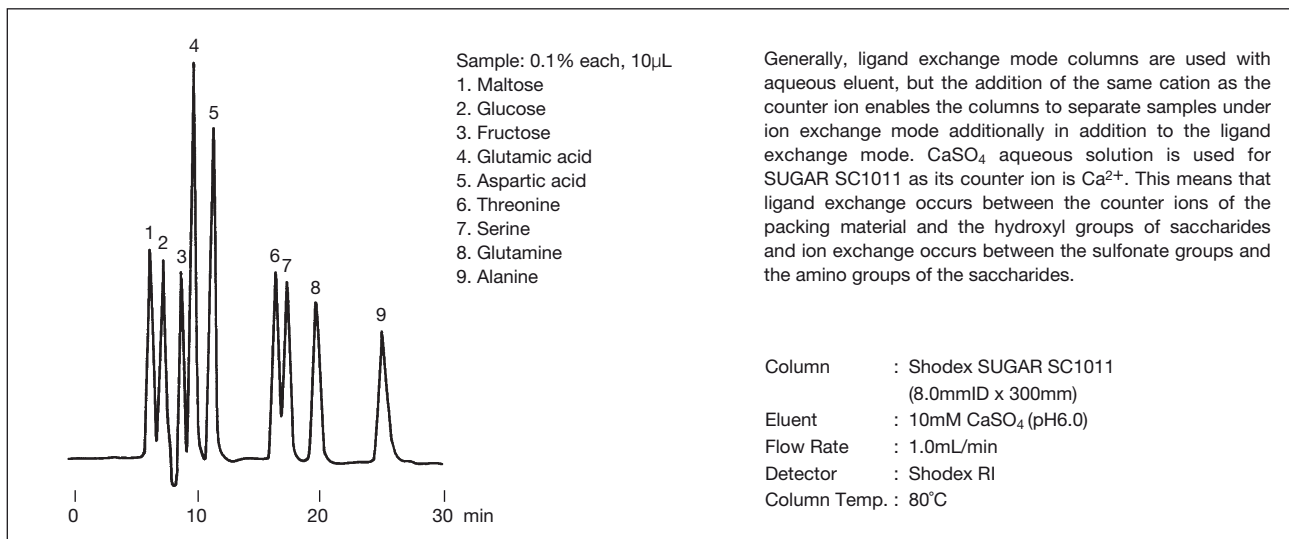


Fig. 2-21 Saccharides and Amino Acids (1)

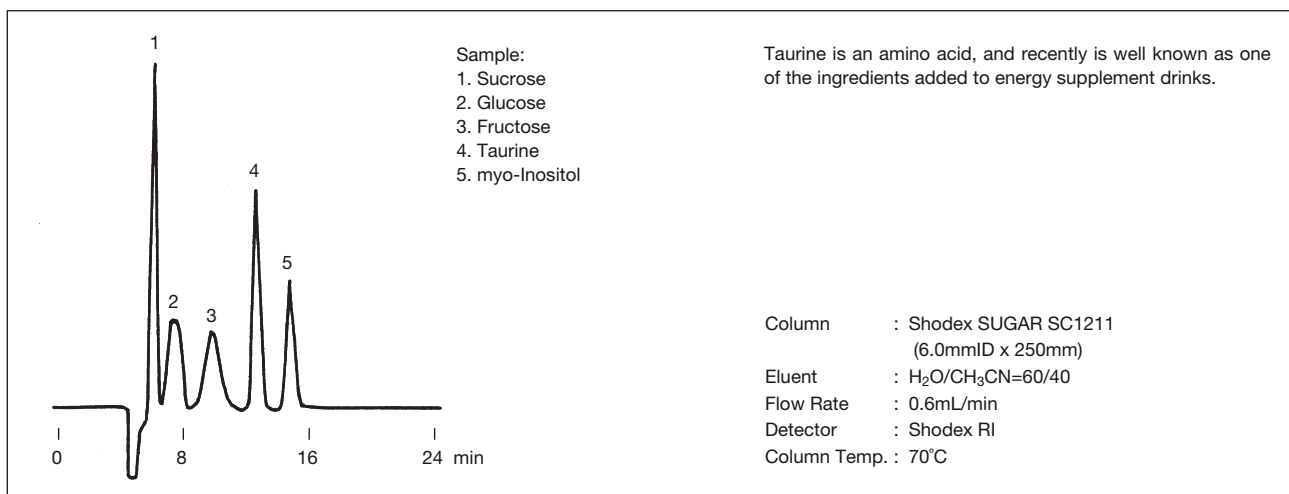


Fig. 2-22 Saccharides and Amino Acids (2)

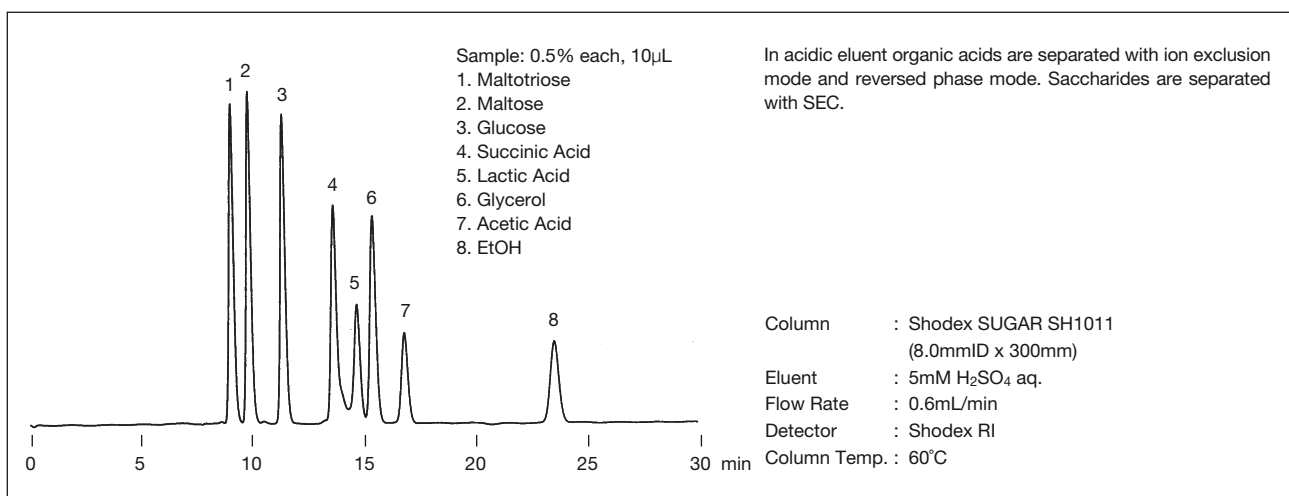


Fig. 2-23 Saccharides and Organic Acids

## 2-6. Polysaccharides

Polysaccharides are used as thickeners/stabilizers to thicken and/or solidify foods and for other purposes. Polysaccharides have a broad range of molecular weights, therefore their analyses are performed not only for quantification but also for molecular weight determination.

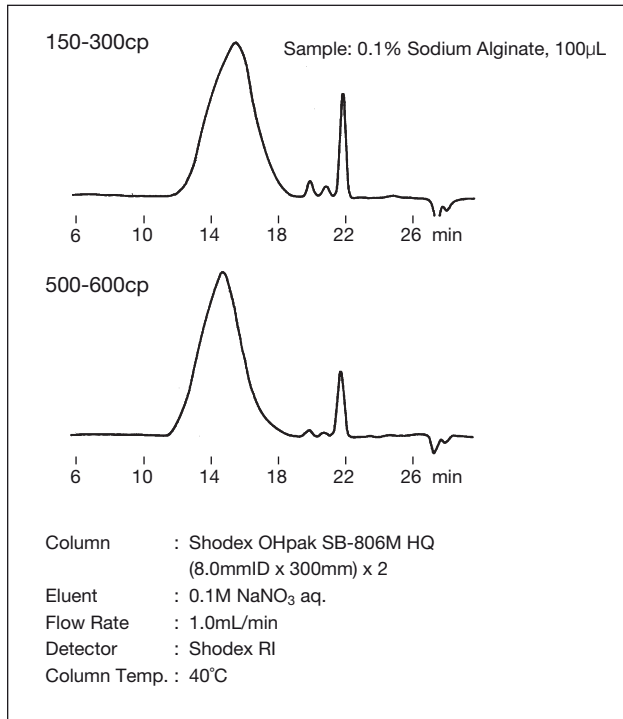


Fig. 2-24 Sodium Alginate

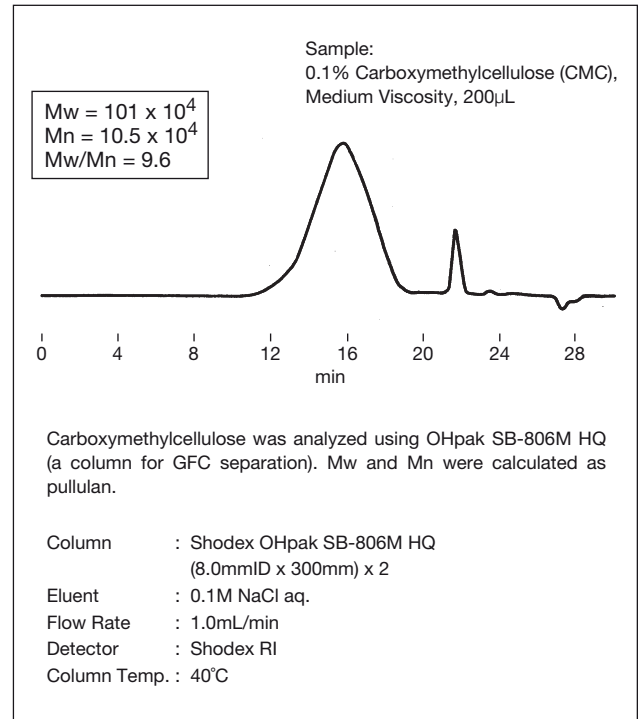


Fig. 2-25 Carboxymethylcellulose

## 2-7. Sweeteners

Generally food additives are analyzed with reversed phase mode columns: however, saccharides are difficult to analyze because of their high polarity. So columns for saccharide analysis are recommended to analyze saccharides and sweeteners at the same time.

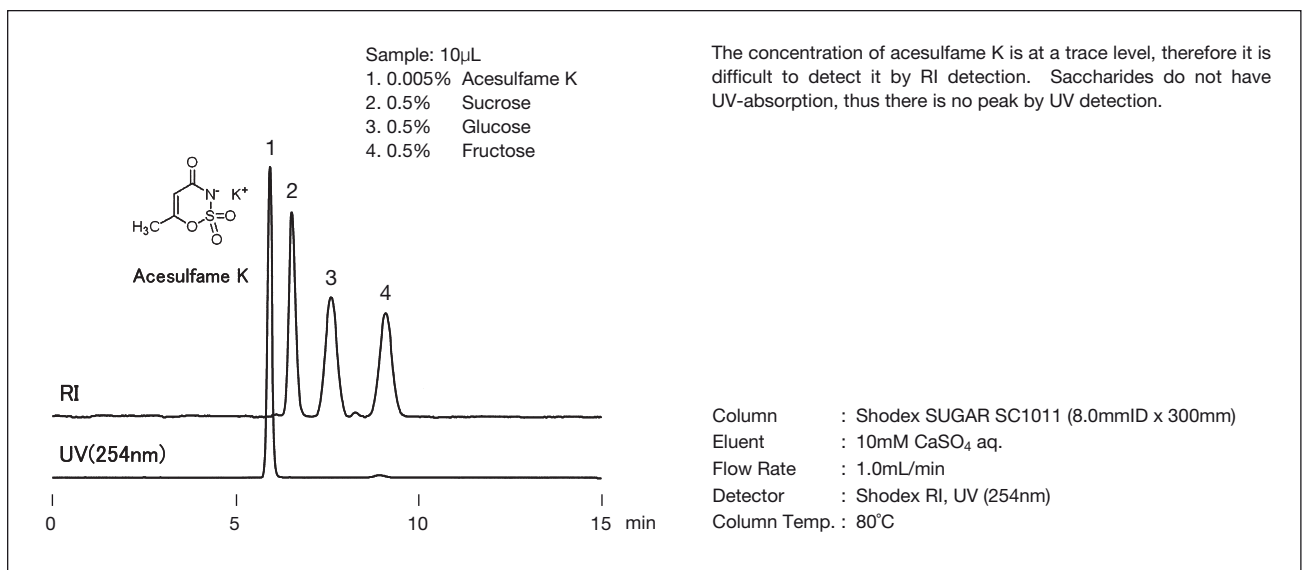


Fig. 2-26 Acesulfame K and Saccharides

Tables of elution volume of saccharides with each column are shown in Appendix-A.

### 3. Organic Acids

Organic acids are weakly acidic substances having carboxyl groups in their molecular structure, and are also referred to as carboxylic acids. When analyzing organic acids, ion exclusion, reversed-phase, ion exchange (ion chromatography), size exclusion, and other types of chromatography are used. Shodex columns for organic acid analysis include RSpak KC-811 (a combination of ion exclusion mode and reversed-phase mode), RSpak DE-413 (polymeric reversed-phase column), and IC SI-50 4E (anion chromatography column), as well as SUGAR SH1011 (column for analysis of saccharides and organic acids, described in Section 2). Table 3-1 shows a lineup of Shodex packing columns for organic acid analysis.

**Table 3-1. Shodex Columns for Organic Acid Analysis**

Product Code	Product Name	Separation Mode*1	Theoretical Plate Number (TP/column)	Particle Size (μm)	ID x Length (mm)
F6378030	RSpak KC-811	IEX+RP	≥17,000	6	8.0 x 300
F6700030	RSpak KC-G	IEX+RP	(Guard column)	10	6.0 x 50
F6700010	RSpak KC-LG	IEX+RP	(Guard column)	12	8.0 x 50
F7008140	RSpak NN-814	IEX+RP	≥9,000	10	8.0 x 250
F6700510	RSpak NN-G	IEX+RP	(Guard column)	10	6.0 x 50
F7001004	RSpak DE-613	RP	≥7,000	6	6.0 x 150
F7001005	RSpak DE-413	RP	≥11,000	4	4.6 x 150
F7009030	RSpak DE-413L	RP	≥17,000	4	4.6 x 250
F7001006	RSpak DE-413S	RP	≥3,000	4	4.6 x 50
F6700150	RSpak DE-G	RP	(Guard column)	10	4.6 x 10
F7001007	RSpak DE-213	RP	≥8,000	4	2.0 x 150
F6700151	RSpak DE-SG	RP	(Guard column)	4	2.0 x 10
F6378100	SUGAR SH1011	SEC + IEX	≥17,000	6	8.0 x 300
F6378101	SUGAR SH1821	SEC + IEX	≥17,000	6	8.0 x 300
F6700080	SUGAR SH-G	-	(Guard column)	10	6.0 x 50
F6995244	IC SI-90 4E	AEC	≥5,000	9	4.0 x 250
F6995245	IC SI-50 4E	AEC	≥10,000	5	4.0 x 250
F6709620	IC SI-90G	AEC	(Guard column)	9	4.6 x 10

\*1 : SEC (Size Exclusion), IEX (Ion Exclusion), RP (Reversed Phase), AEC (Anion Exchange)

#### 3-1. Separation Mechanism

Shodex can provide several columns for analysis of organic acids depending on other substances in the sample. (Table 3-2)

**Table 3-2. Column Selection for Samples**

Column	Separation Mode	Sample
RSpak KC-811	Ion Exclusion + Reversed Phase	General Organic Acids
RSpak NN-814	Ion Exclusion + Reversed Phase	Aromatic Organic Acids
RSpak DE-413	Reversed Phase	General Organic Acids
SUGAR SH1011 SUGAR SH1821	Size Exclusion + Ion Exclusion	Organic Acids and Sugars
IC SI-90 4E IC SI-50 4E	Anion Exchange	Organic Acids and Anions

### 3-1-1. Ion Exclusion Mode

In organic acid analysis, a strong cation exchange resin with sulfo-groups bound to the packing surface is used. Weak acids, such as organic acids, exhibit only partial dissociation when dissolved in water. In a dissociated state (negatively charged), the positive charge of the cation is neutralized and excluded by the negative charge of the sulfo-groups on the resin surface. There is no concern over organic acid adsorption into the resin. In a non-dissociated state, on the other hand, ion exclusion does not occur; hydrophobic adsorption into the resin substrate occurs instead to allow organic acids to be retained by the resin (Figure 3-1).

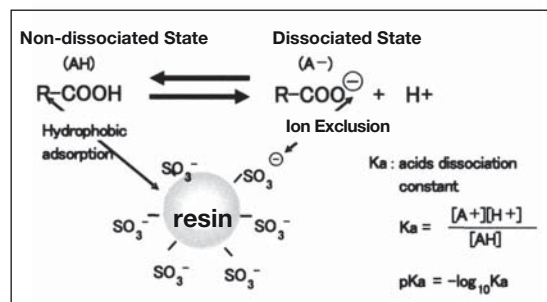


Fig. 3-1 Ion Exclusion Mode

Hence, organic acids are eluted in ascending order of pKa (descending order of acidity) and in descending order of polarity. The packing substrate of KC-811 is a styrene-divinylbenzene copolymer, therefore it is recommended to use NN-814, which has a higher polarity of packing substrate than KC-811, when analyzing aromatic organic acids of low polarity. The packing substrate of NN-814 is poly(hydroxymethacrylate).

### 3-1-2. Reversed Phase Mode

The reversed-phase mode is the most commonly used separation mode of HPLC. This mode is based on the hydrophobic interaction between the portion of low polarity of the packing and the portion of low polarity of the sample. Hence, elution occurs in descending order of polarity. A key to successful organic acid analysis in reversed-phase mode resides in thoroughly suppressing the dissociation of organic acids and reducing the polarity of the sample. Organic acids are generally reported to become non-dissociated when the eluent has a pH value lower than the pKa by 1.5.

### 3-1-3. Anion Exchange Mode

The resin surface is positively charged. The cation on the resin is scrambled by the anion of the component mobile phase and the organic acid anion; elution occurs in ascending order of ionic adsorptivity.

### 3-1-4. Size Exclusion Mode

Organic acids are eluted in ascending order of molecular size. Molecules larger than the pore size of resin are eluted  $V_0$  position.

### 3-1-5. Detection of Organic Acids

The functional group shared by organic acids is the carboxyl group, which has a UV absorption band between 200 and 210nm, thus enabling the use of a UV detector. However, because many other organic substances have the same wavelength band for UV absorption, and also because the molar extinction coefficient is low at 50 to 70, detection of organic acids is likely to be influenced by impurities. The RI detector lacks selectivity and remains somewhat problematic with regard to sensitivity. Detection using the conductivity detector (CD) is prone to sensitivity variation depending on the kind of organic acid. As a method of selectively detecting organic acids, the post-column method using a pH indicator is available which employs a visible absorption spectrum (VIS) detector. (Fig. 3-2)

Column	: Shodex RSpak KC-G (6.0mmID x 50mm) + KC-811 (8.0mmID x 300mm)x2	Sample:
Eluent	: 3mM HClO <sub>4</sub> aq.	1. 800ppm Citric Acid
Flow Rate	: 1.0mL/min	2. 900ppm Tartaric Acid
Column Temp.	: 50°C	3. 800ppm Malic Acid
		4. 700ppm Succinic Acid
		5. 1100ppm Lactic Acid
		6. 600ppm Formic Acid
		7. 700ppm Acetic Acid
		8. 1500ppm Levulinic Acid
		9. 1500ppm Pyroglutamic Acid

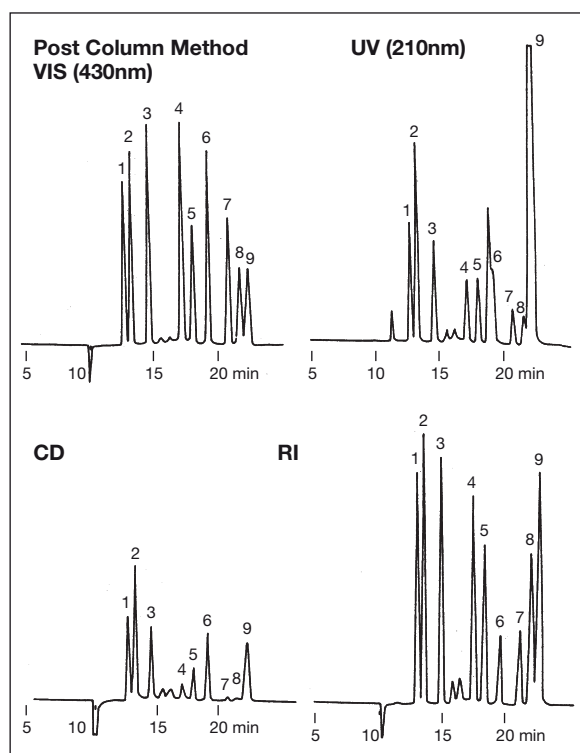


Fig. 3-2 Comparison of Chromatograms with Various Detectors



### 3-2. Organic Acids

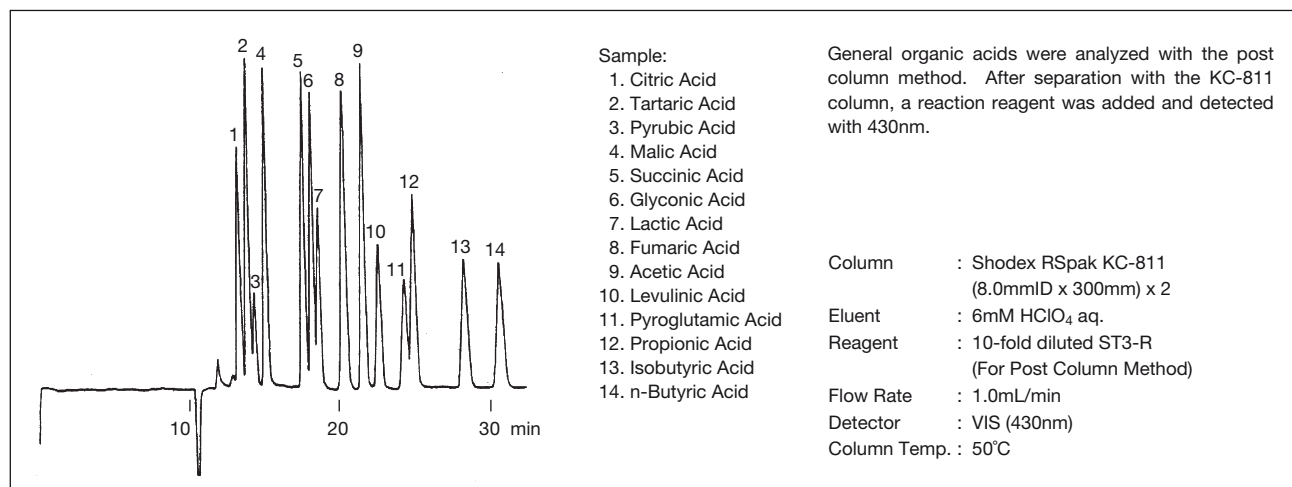


Fig. 3-3. Analysis of Organic Acids with Post Column Method

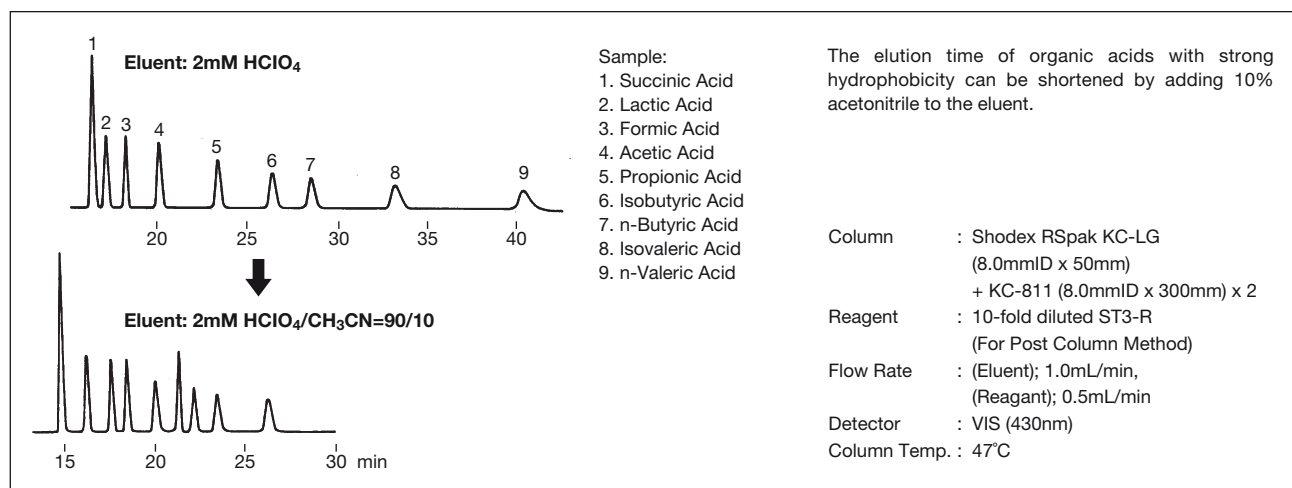


Fig. 3-4 Effect of Elution of Organic Acids with Addition of Acetonitrile

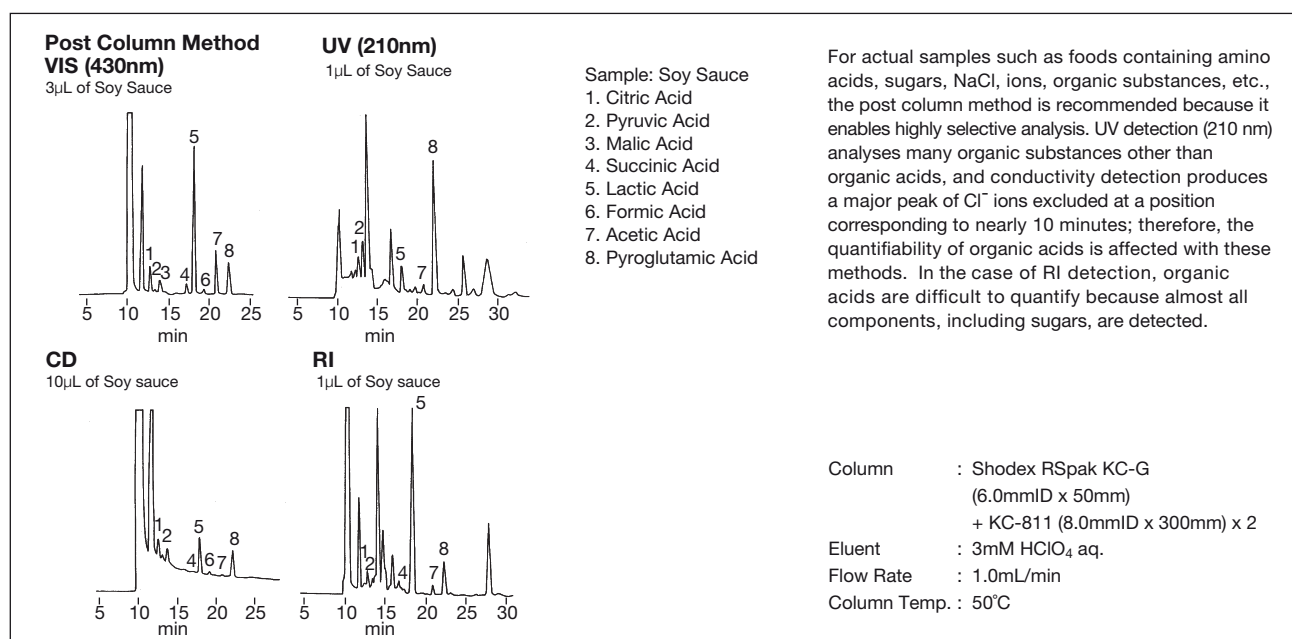


Fig. 3-5 Comparison of Results of Soy Sauce with Various Detectors

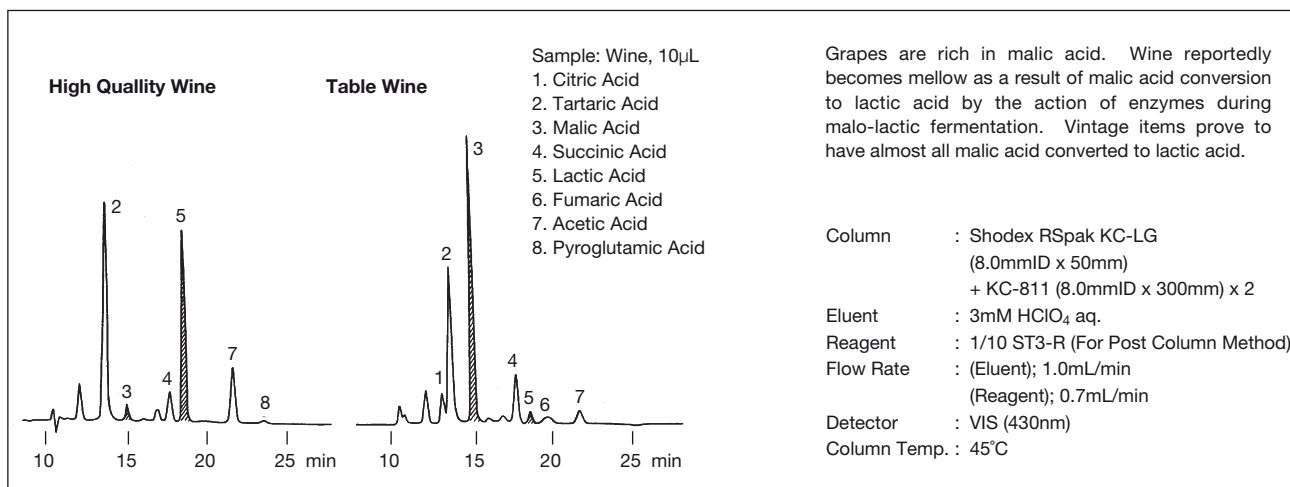


Fig. 3-6 Organic Acids in White Wine

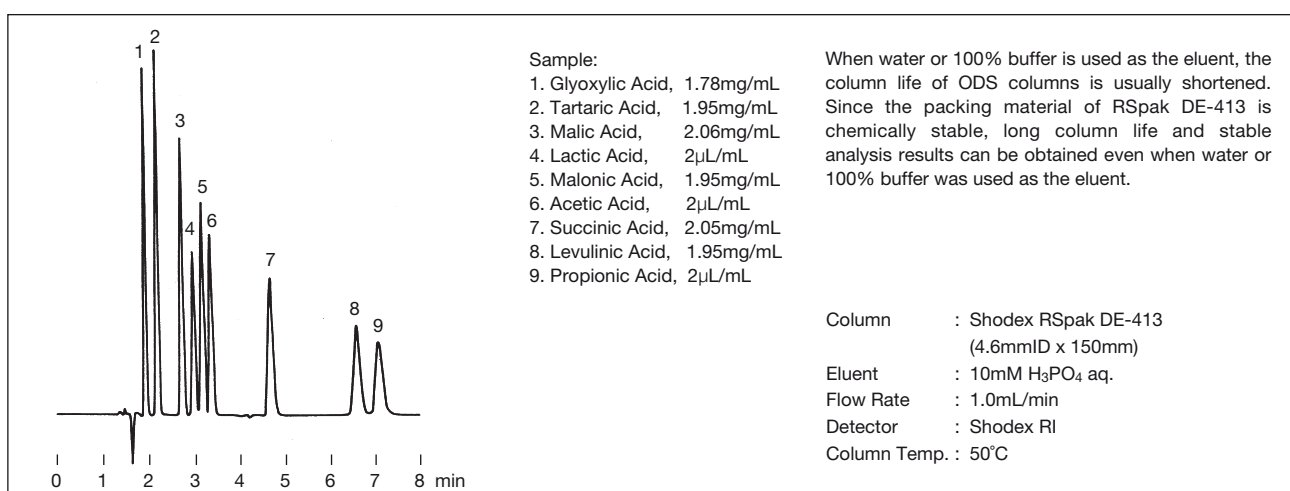


Fig. 3-7 Organic Acids (Reversed Phase Mode)

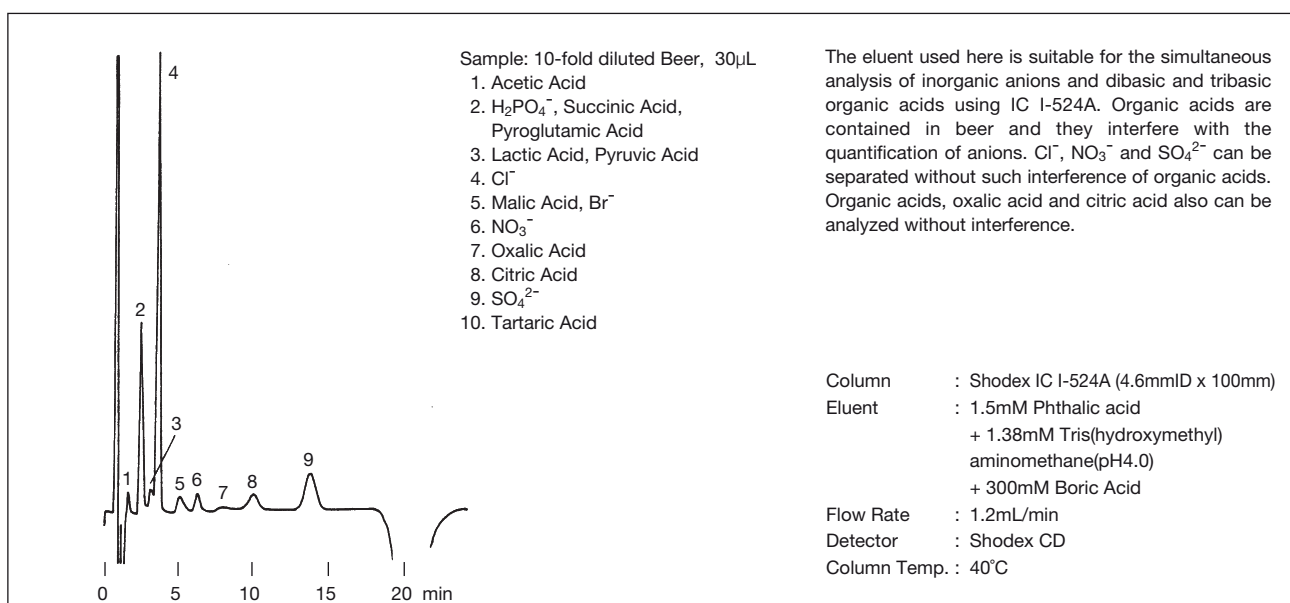


Fig. 3-8 Organic Acids and Anions in Beer (Anion Exchange Mode)

Tables of elution volume of organic acids with each column are shown in appendix-B, C and D.

## 4. Vitamins

There are two types of vitamins: fat-soluble vitamins and water-soluble vitamins. (Fig. 4-1)

It is possible to analyze vitamins simultaneously, though several official methods show the analysis methods respectively. The major analysis methods by HPLC are reversed phase or normal phase, additionally size exclusion chromatography can be applied. Table 4-1 shows the lineup of Shodex packed columns for vitamin analysis with separation modes.

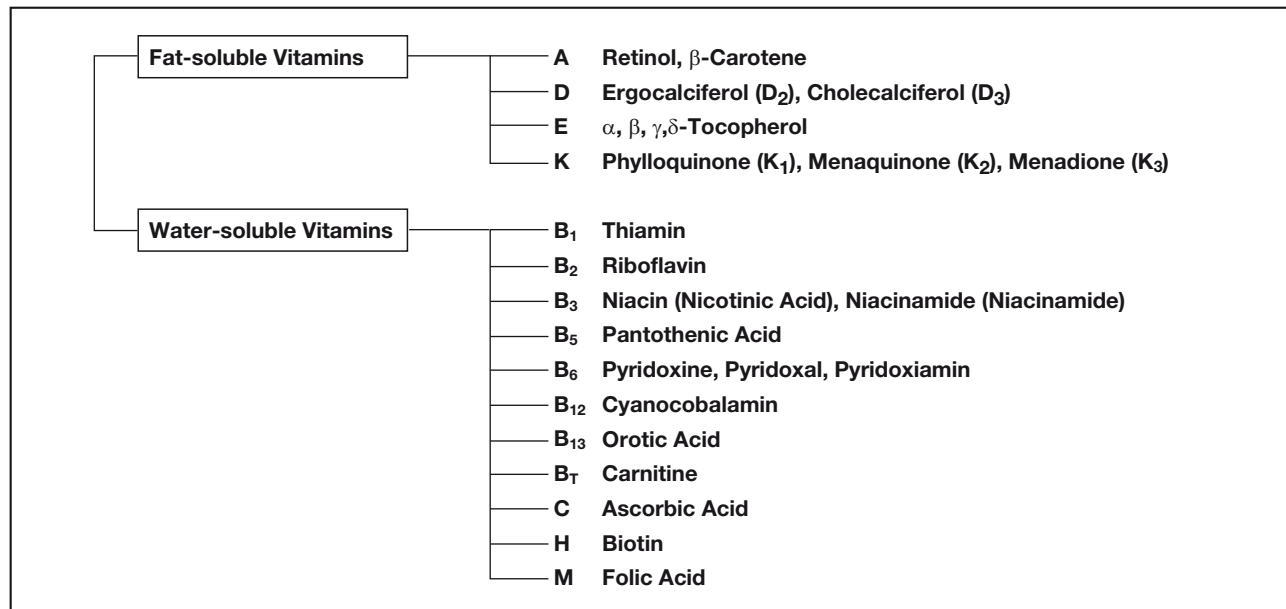


Fig. 4-1 Categories of Vitamins

Table 4-1 Shodex Columns for Vitamin Analysis  
(Absorption and Partition)

Product Code	Product Name	Theoretical Plate Number (TP/column)	Particle Size (μm)	ID x Length (mm)
F7621001	Asahipak ODP-40 4D	≥11,000	4	4.6 x 150
F7621002	Asahipak ODP-40 4E	≥17,000	4	4.6 x 250
F7620002	Asahipak ODP-50 6D	≥ 9,000	5	6.0 x 150
F7620001	Asahipak ODP-50 6E	≥14,000	5	6.0 x 250
F6710001	Asahipak ODP-50G 6A	(Guard column)	5	6.0 x 10
F6710023	Asahipak ODP-50 4B	≥2,500	5	4.6 x 50
F7620004	Asahipak ODP-50 4D	≥9,000	5	4.6 x 150
F7620003	Asahipak ODP-50 4E	≥14,000	5	4.6 x 250
F6710022	Asahipak ODP-50G 4A	(Guard column)	5	4.6 x 10
F7620009	Asahipak ODP-50 2D	≥5,000	5	2.0 x 150
F6713001	Asahipak ODP-50G 2A	(Guard column)	5	2.0 x 10
F7001004	RSpak DE-613	≥7,000	6	6.0 x 150
F7001005	RSpak DE-413	≥11,000	4	4.6 x 150
F7009030	RSpak DE-413L	≥17,000	4	4.6 x 250
F7001006	RSpak DE-413S	≥3,000	4	4.6 x 50
F6700150	RSpak DE-G	(Guard column)	10	4.6 x 10
F7001007	RSpak DE-213	≥8,000	4	2.0 x 150
F6700151	RSpak DE-SG	(Guard column)	4	2.0 x 10
F7630002	Asahipak NH2P-50 4D	≥5,500	5	4.6 x 150
F7630001	Asahipak NH2P-50 4E	≥7,500	5	4.6 x 250
F6710016	Asahipak NH2P-50G 4A	(Guard column)	5	4.6 x 10
F7630006	Asahipak NH2P-50 2D	≥3,500	5	2.0 x 150
F6713000	Asahipak NH2P-50G 2A	(Guard column)	5	2.0 x 10
F6650050	Silica 5SIL 4D	≥9,000	5	4.6 x 150
F6650051	Silica 5SIL 4E	≥15,000	5	4.6 x 250

( Size Exclusion )

Product Code	Product Name	Theoretical Plate Number (TP/column)	Particle Size (μm)	Exclusion Limit (Polystyrene)	ID x Length (mm)
F6028010	GPC KF-801	≥18,000	6	1,500	8.0 x 300
F6700300	GPC KF-G	(Guard column)	8		4.6 x 10

( Multimode )

Product Code	Product Name	Theoretical Plate Number (TP/column)	Particle Size (μm)	ID x Length (mm)
F7008140	RSpak NN-814	≥9,000	10	8.0 x 250
F7008150	RSpak NN-614	≥4,000	10	6.0 x 150
F6700510	RSpak NN-G	(Guard column)	10	6.0 x 50
F7008160	RSpak NN-414	≥6,000	10	4.6 x 150

#### 4-1. Fat-Soluble Vitamins

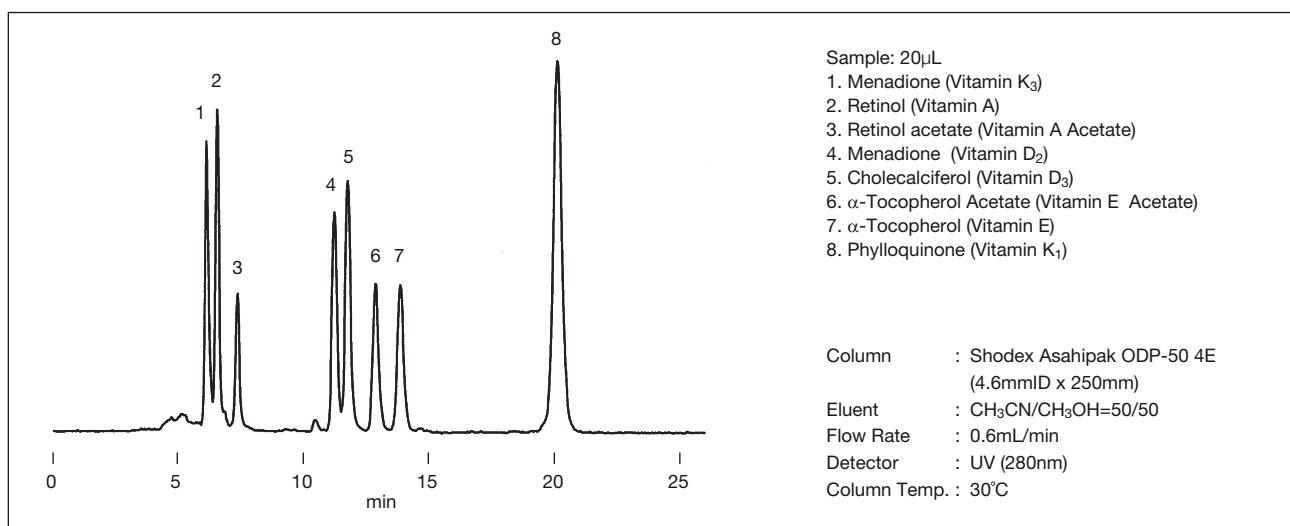


Fig. 4-2 Simultaneous Analysis of Fat-soluble Vitamins (Reversed Phase Mode)

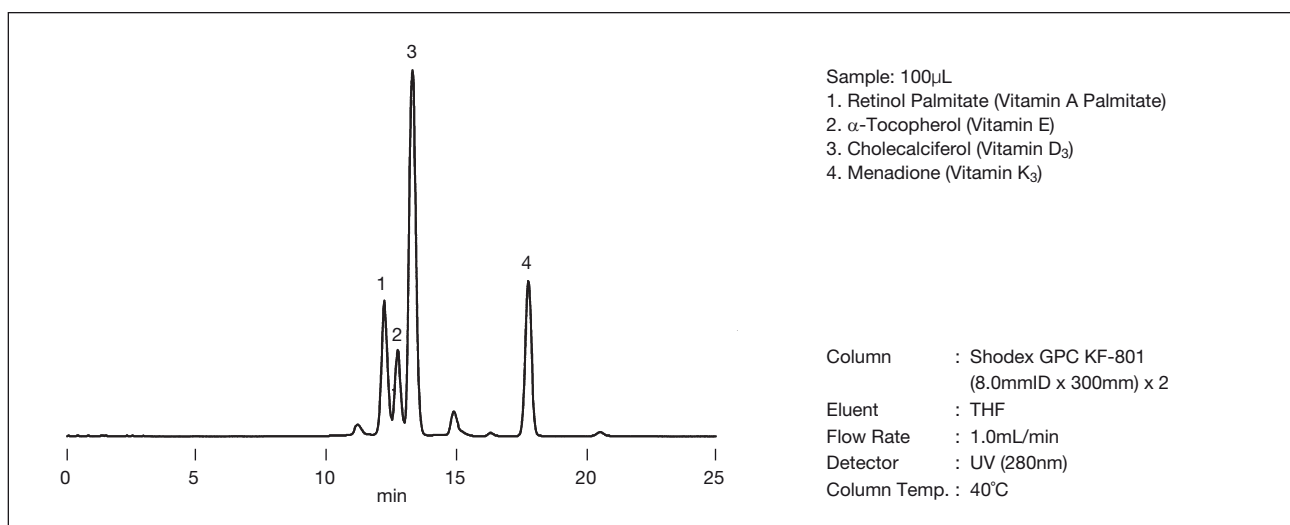


Fig. 4-3 Simultaneous Analysis of Fat-soluble Vitamins (Size Exclusion Mode)

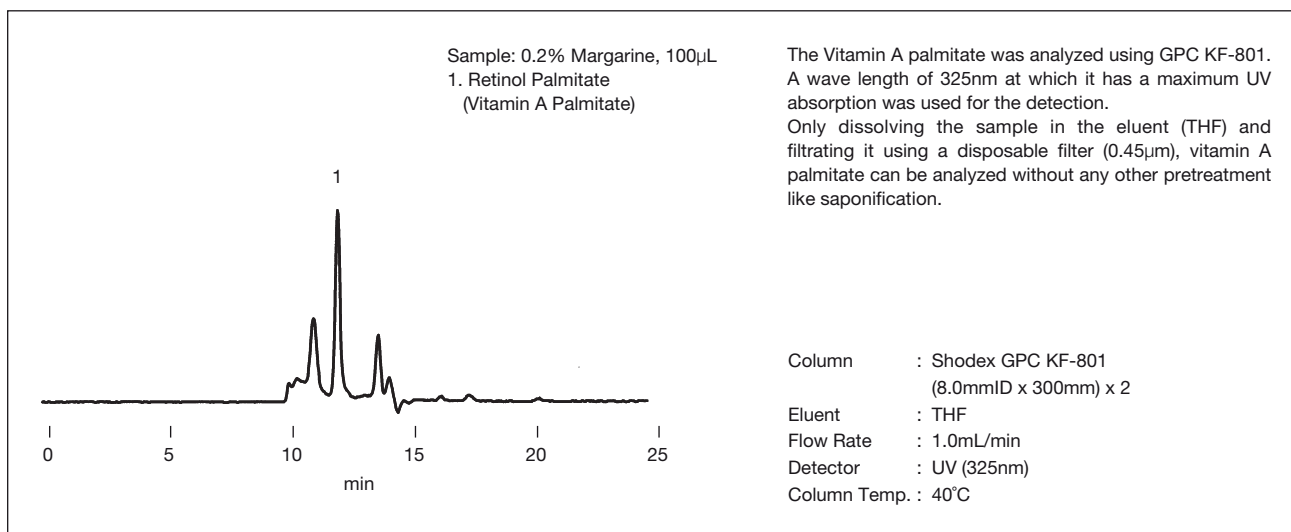


Fig. 4-4 Vitamin A Palmitate in Margarine (Size Exclusion Mode)

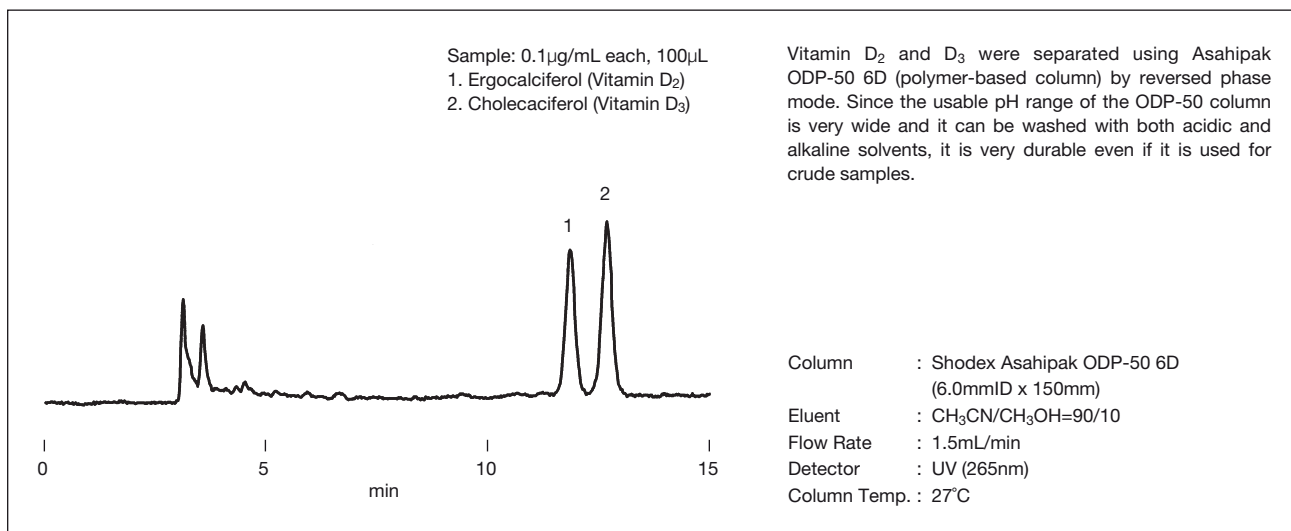


Fig. 4-5 Vitamin D<sub>2</sub> and D<sub>3</sub> (Reversed Phase Mode)

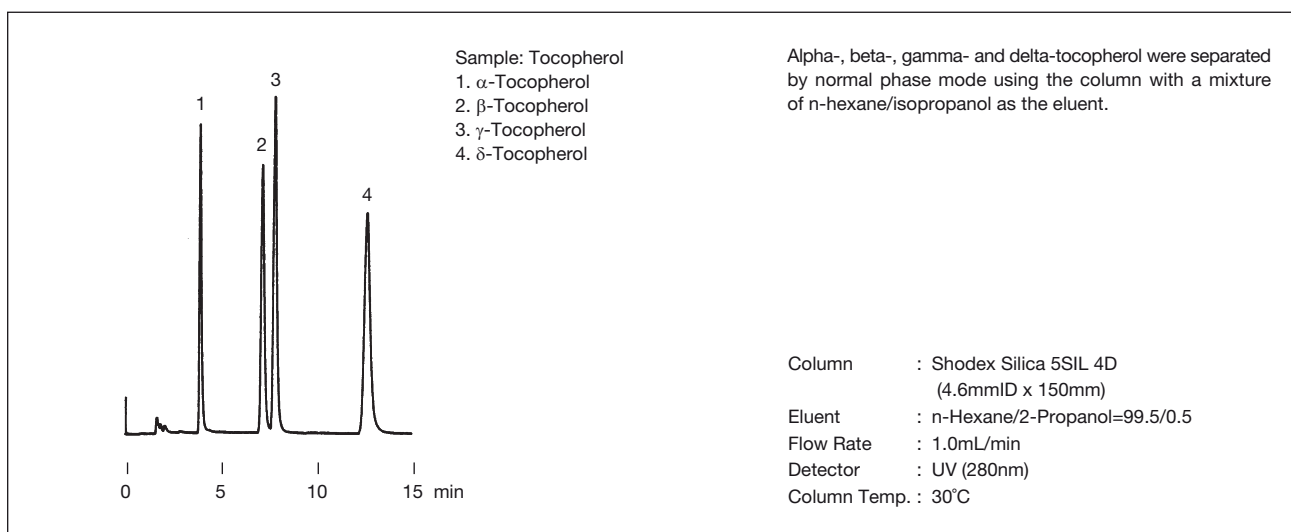


Fig. 4-6 Vitamin E (Normal Phase Mode)

## 4-2. Water-soluble Vitamins

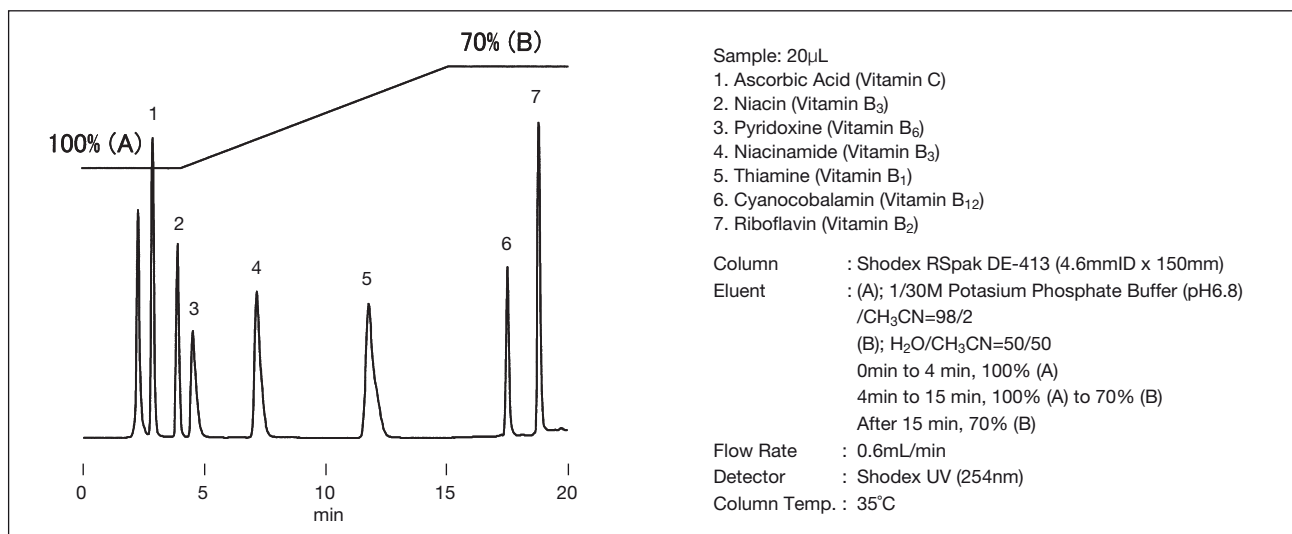


Fig. 4-7 Simultaneous Analysis of Water-soluble Vitamins (Reversed Phase Mode)

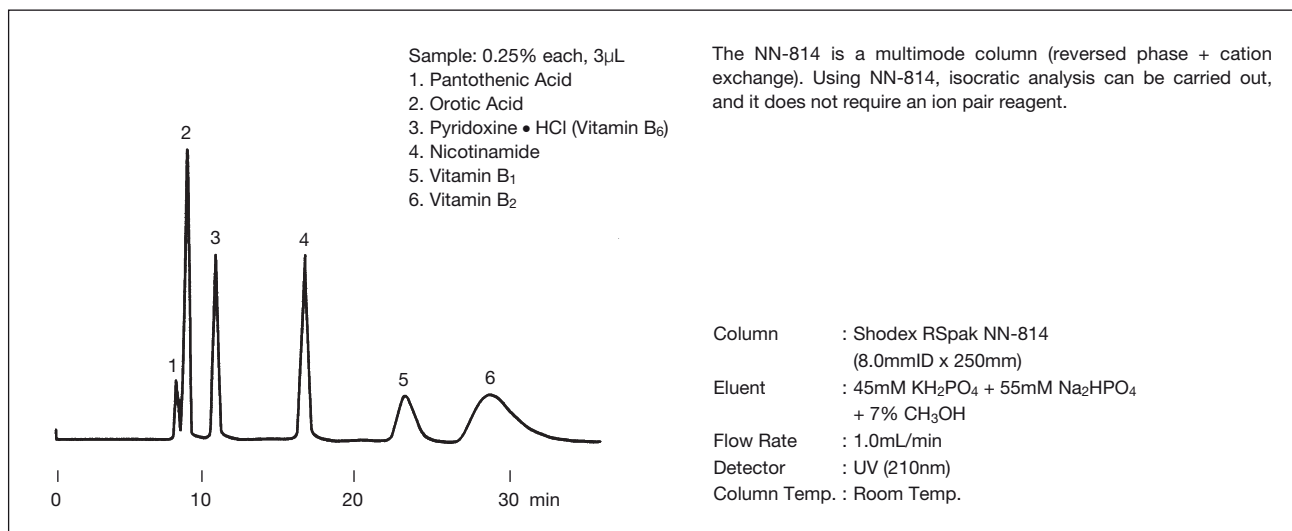


Fig. 4-8 Simultaneous Analysis of Water-soluble Vitamins (Multimode)

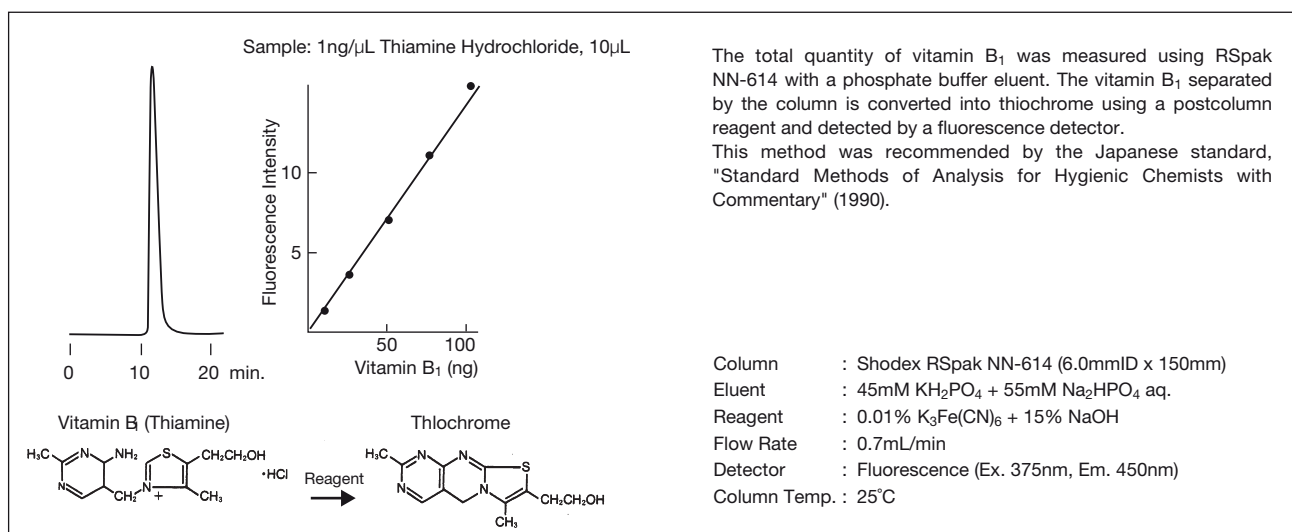


Fig. 4-9 Vitamin B1 (Multimode)

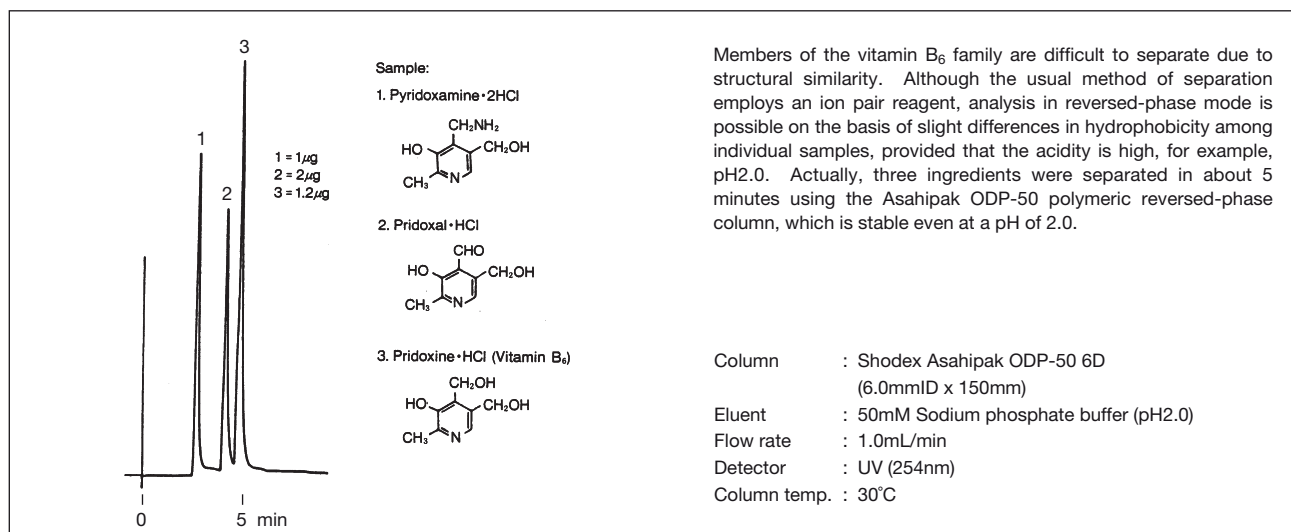


Fig. 4-10 Vitamin B<sub>6</sub> (reversed phase mode)

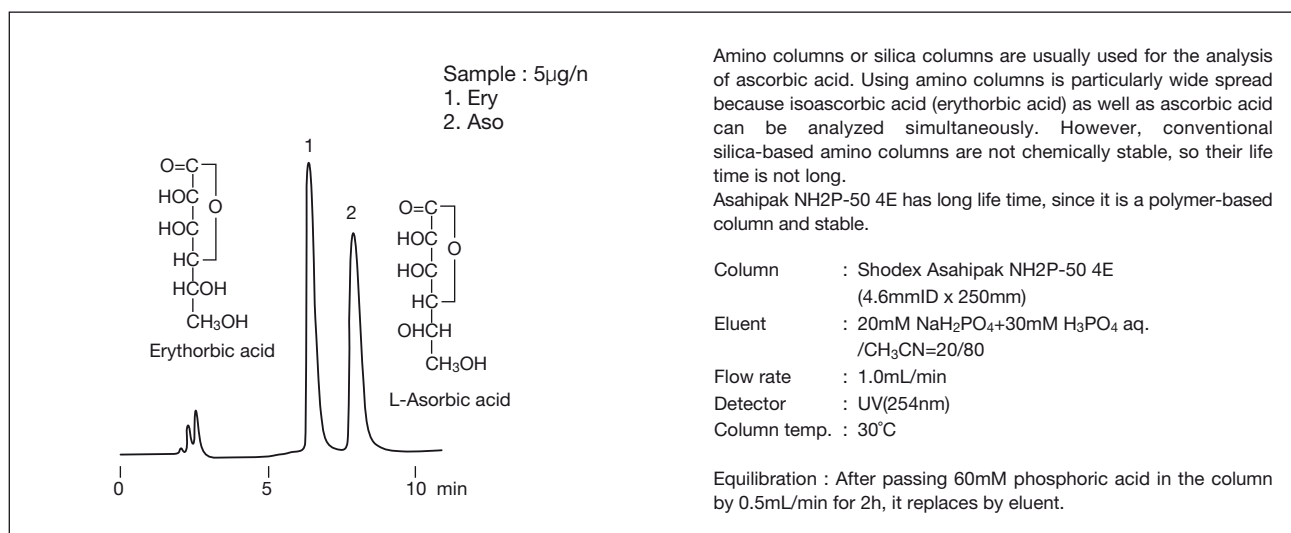


Fig. 4-11 Vitamin C (normal phase mode)

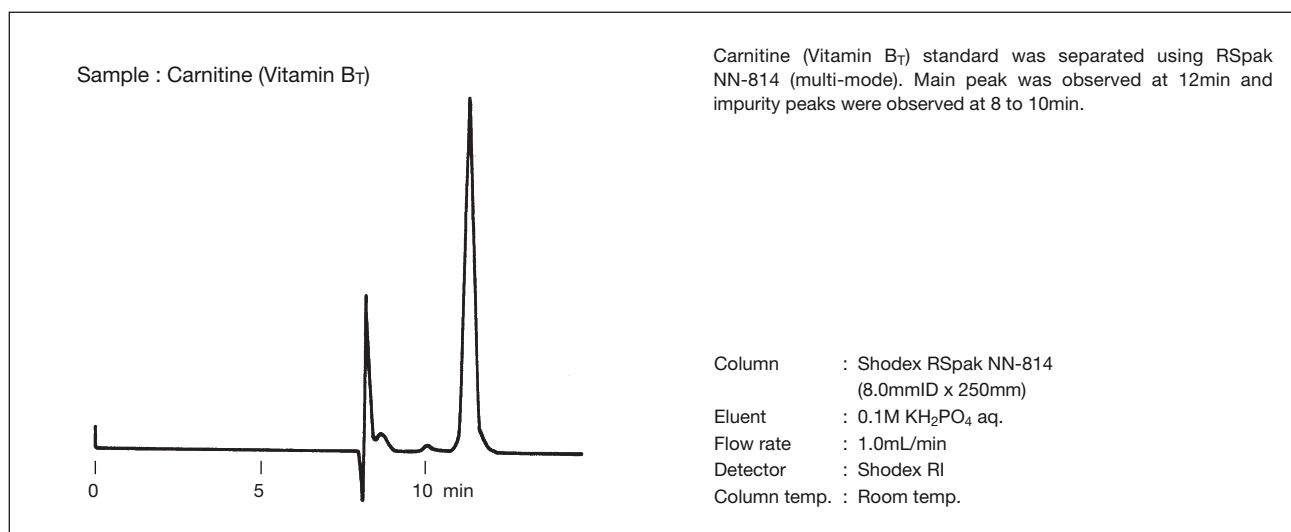


Fig. 4-12 Vitamin B<sub>7</sub> (multi- mode)



## 5. Other Ingredients in Foods

### 5-1. Higher Fatty Acids

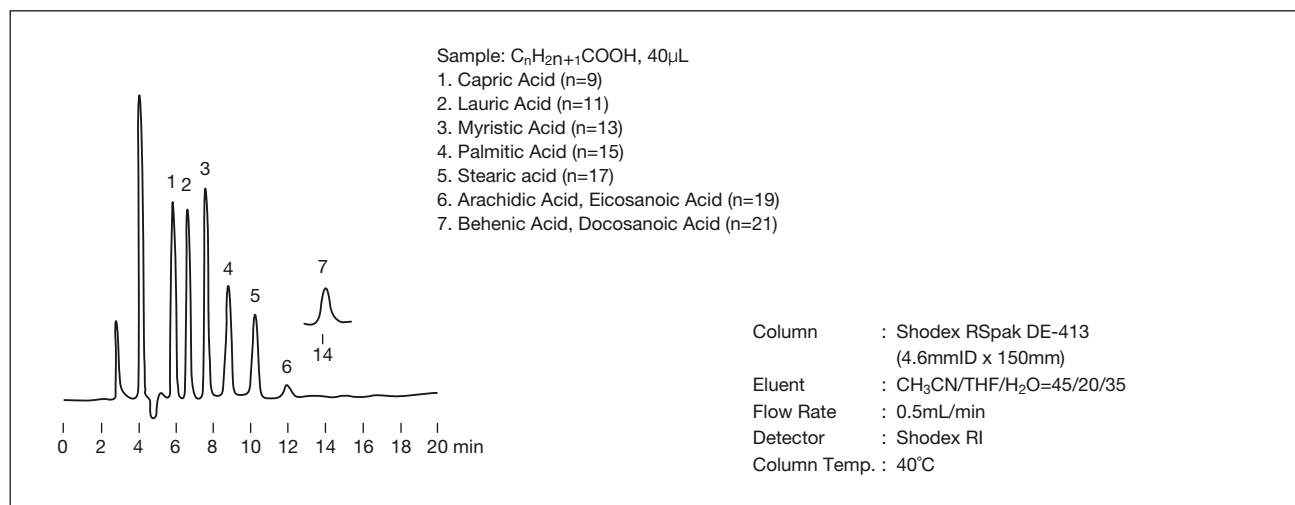


Fig. 5-1 Higher Fatty Acids (1)

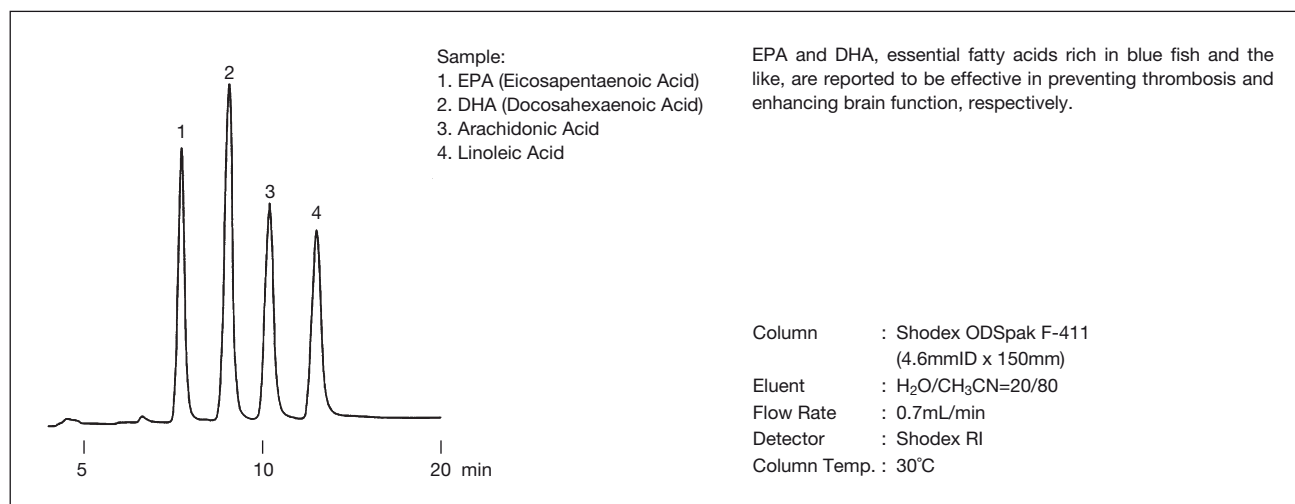


Fig. 5-2 Higher Fatty Acids (2)

Table 5-1 Shodex Columns for Higher Fatty Acid Analysis (Absorption and Partition)

Product Code	Product Name	Theoretical Plate Number (TP/column)	Particle Size ( $\mu$ m)	Carbon Content (%)	ID x Length (mm)
F7001004	RSpak DE-613	$\geq 7,000$	6	—	6.0 x 150
F7001005	RSpak DE-413	$\geq 11,000$	4	—	4.6 x 150
F7009030	RSpak DE-413L	$\geq 17,000$	4	—	4.6 x 250
F7001006	RSpak DE-413S	$\geq 3,000$	4	—	4.6 x 50
F6700150	RSpak DE-G	(Guard column)	10	—	4.6 x 10
F7001007	RSpak DE-213	$\geq 8,000$	4	—	2.0 x 150
F6700151	RSpak DE-SG	(Guard column)	4	—	2.0 x 10
F6604112	ODSpak F-411	$\geq 8,000$	5	14	4.6 x 150
F6605110	ODSpak F-511	$\geq 14,000$	5	14	4.6 x 250
F6604113	ODSpak F-411/S	$\geq 10,000$	3	14	4.6 x 100

## 5-2. Nucleic Acids as Taste Components

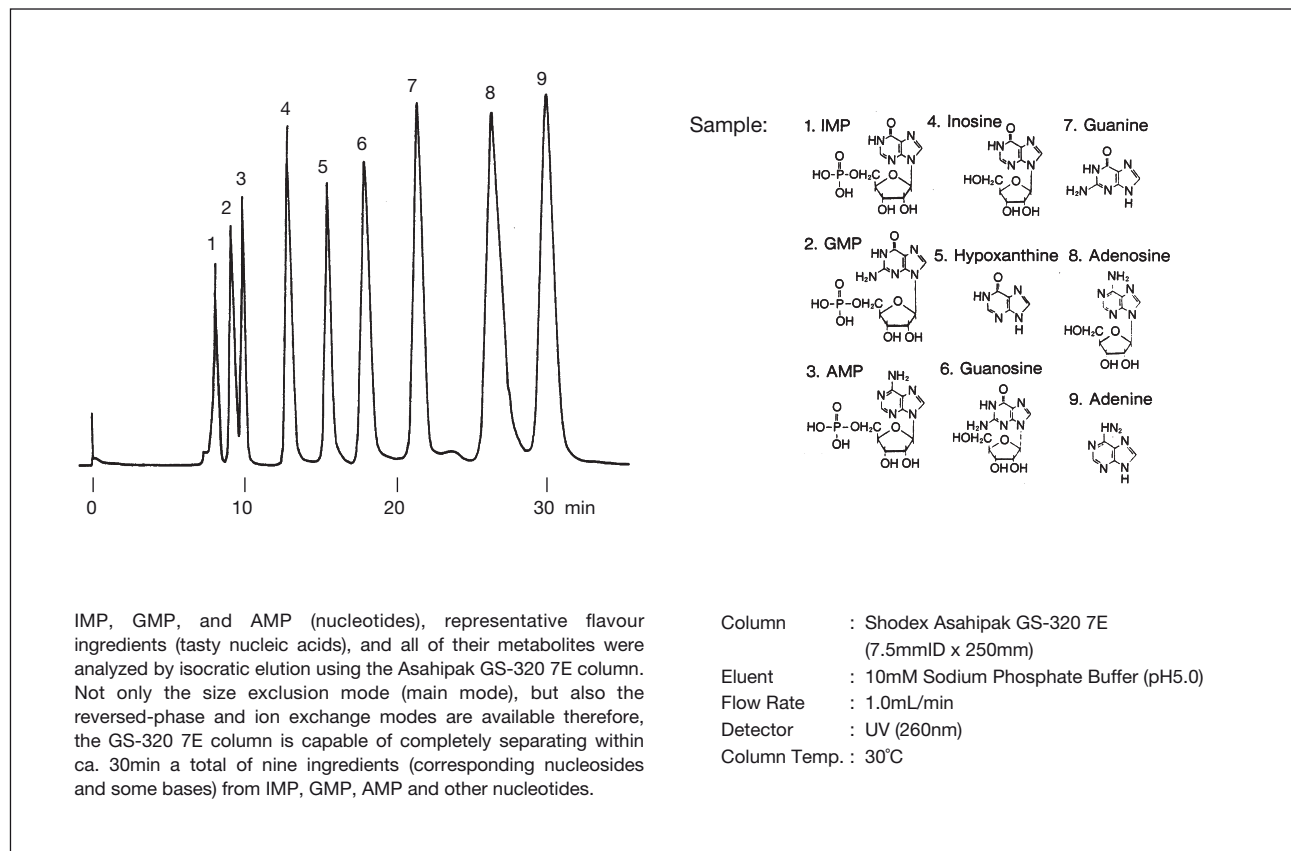


Fig. 5-3 Nucleic Acids as Taste Components

Table 5-2 Shodex Columns for Flavour Compounds Analysis (Multimode)

Product Code	Product Name	ID x Length (mm)
F7610005	Asahipak GS-320 7E	7.5 x 300
F6710019	Asahipak GS-2G 7B	7.5 x 50

### 5-3. Amino Acids

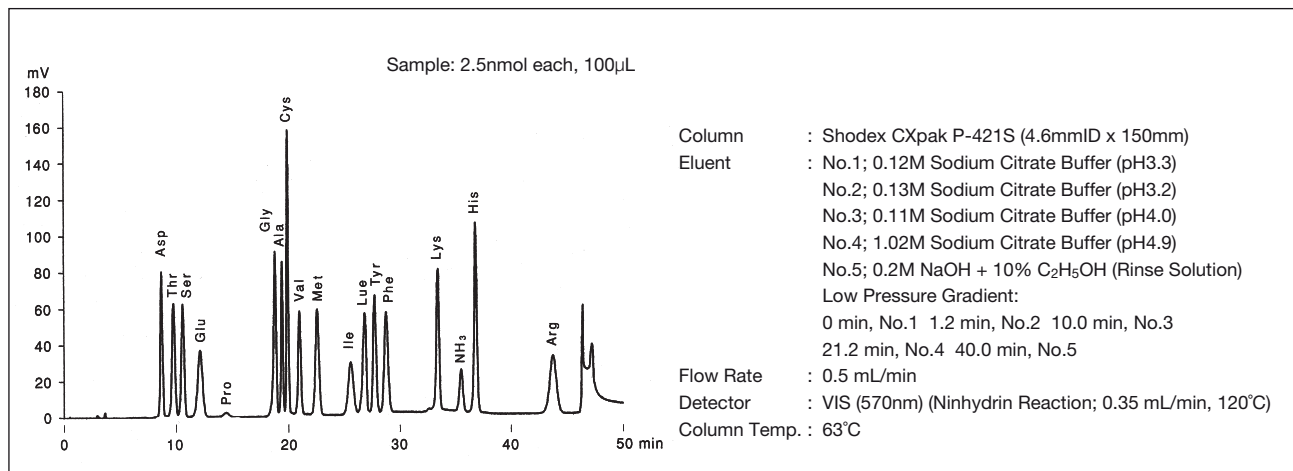


Fig. 5-4 Amino Acids (Strong Cation Exchange Mode)

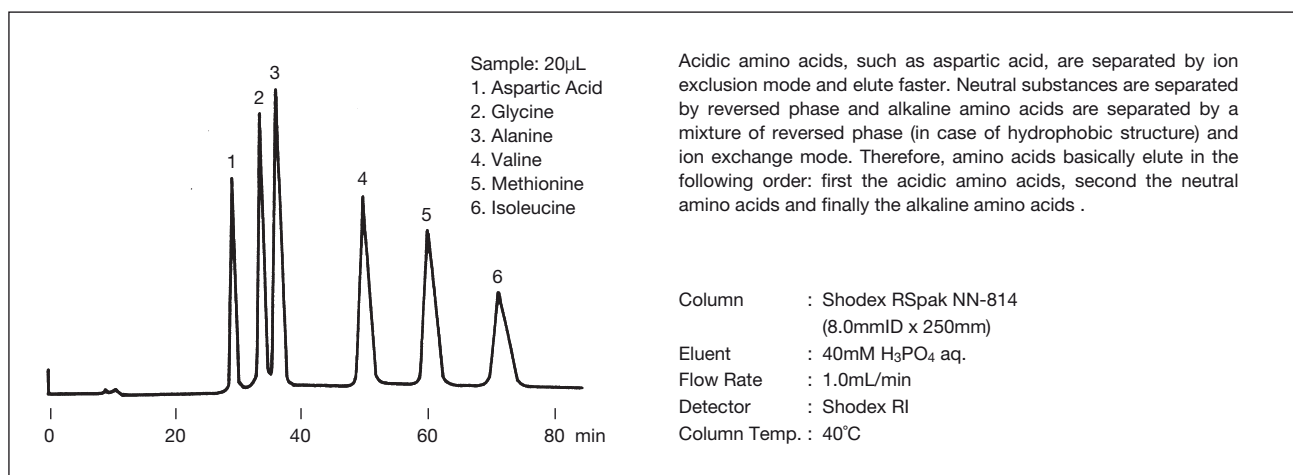


Fig. 5-5 Amino Acids (Multimode)

Table 5-3 Shodex Columns for Amino Acids Analysis (Strong Cation Exchange)

Product Code	Product Name	SeparationMode	Particle Size (µm)	ID x Length (mm)
F6354211	CXpak P-421S	Strong Cation Exchange	6	4.6 x 150
F6700210	CXpak P-G		10	4.6 x 10

Table 5-3 Shodex Columns for Amino Acids Analysis (Multimode)

Product Code	Product Name	SeparationMode	Particle Size (µm)	ID x Length (mm)
F7008140	RSpak NN-814	Ion Exclusion	10	8.0 x 250
F7008150	RSpak NN-614	+ Ion Exchange	10	6.0 x 150
F6700510	RSpak NN-G	+ Reversed Phase	10	6.0 x 50
F7008160	RSpak NN-414		10	4.6 x 150

## Appendix

### A. List of Elution volume of Saccharides

Substances	Elution Volume (mL)						
	SUGAR Series					Asahipak	Asahipak
	SP0810 Pb <sup>2+</sup>	SC1011 Ca <sup>2+</sup>	KS-801 Na <sup>+</sup>	SZ5532 Zn <sup>2+</sup>	SC1211 Ca <sup>2+</sup>	NH2P-50 4E	GS-220 HQ ×2
N-Acetyl- $\alpha$ -D-glucosamine	8.9	7.8	6.7	-	4.1	6.7	
D(+)-Arabinose	10.4	8.9	8.2	5.1	5.6	6.2	
D-Arabitol	15.9	11.3	7.6	7.3	8.2	6.3	
Aspartame	-	-	-	-	34.0	-	
2-Deoxy-D-glucose	8.8	7.6	7.2	4.3	4.0	6.0	
Difructose Anhydride (II)	7.1	6.3	5.8	4.3	*	7.8	
Dulcitol	20.2	12.8	7.4	9.5	11.3	7.5	
meso-Erythritol	12.7	10.1	7.9	5.7	6.3	5.4	17.4
Ethanol	11.1	11.3	10.1	-	4.3	-	
1-Fructofuranosyl-D-nystose	6.1	5.3	4.8	31.4	*	-	14.2
D(-)-Fructose	11.1	8.9	7.7	5.4	5.9	6.8	17.1
D(+)-Fucose	10.5	8.8	8.1	4.5	5.0	5.4	
D(+)-Galactose	9.7	8.0	7.6	6.5	5.0	8.1	16.6
4'-Galactosyllactose	7.4	6.0	5.4	19.0	*	21.7	
$\alpha$ -D-Galacturonic Acid	-	6.3	4.4	-	5.6	-	
Gentiobiose	7.2	6.1	5.8	10.5	*	16.4	
Glucose	8.6	7.3	7.2	5.9	4.8	8.6	16.8
Glycerol							17.8
Glycyrrhizin	-	-	-	-	2.7	-	
myo-Inositol	12.8	8.9	8.0	12.6	7.9	10.0	
Isomaltose	7.7	6.3	6.0	10.6	*	15.2	15.6
Isomaltotriose	7.1	5.8	5.3	21.2	*	27.6	14.7
1-Kestose	6.8	5.8	5.3	13.1	*	20.1	15.3
Kojibiose	7.6	6.2	5.9	9.7	*	14.8	
Lactitol	13.3	8.1	6.1	16.4	6.7	11.8	15.5
Lactose	8.1	6.5	6.0	10.1	4.1	13.3	15.7
Lactosylfructoside	7.1	5.9	5.3	14.7	*	19.0	
Lactulose	9.1	7.0	6.2	9.2	4.7	10.7	
Maltitol	12.2	8.3	6.0	13.0	6.8	11.8	15.8
Maltoheptaose							14.8
Maltohexaose							14.6
Maltopentaose							14.3
Maltose	7.9	6.3	5.9	8.7	*	14.2	16.1
Maltotriose	7.5	5.9	5.4	13.8	*	25.0	15.6
Mannitol	15.8	11.1	7.2	8.8	9.0	7.4	
D-Mannose	10.7	8.2	7.6	5.8	5.0	7.8	
D(+)-Melezitose	6.9	5.8	5.2	13.6	*	19.3	
Melibiose	8.2	6.5	6.0	11.7	4.2	14.7	
Methyl- $\alpha$ -D-mannopyranoside	11.1	8.9	7.8	4.0	4.4	4.7	
Nystose	6.4	5.5	4.9	20.1	*	31.9	14.7
Palatinit	2peaks	2peaks	5.9	2peaks	2peaks	12.7	

(-) → cannot be detected (\*) → cannot be separated from solvent peak

Substances	Elution Volume (mL)						
	SUGAR Series					Asahipak	Asahipak
	SP0810 Pb <sup>2+</sup>	SC1011 Ca <sup>2+</sup>	KS-801 Na <sup>+</sup>	SZ5532 Zn <sup>2+</sup>	SC1211 Ca <sup>2+</sup>	NH2P-50 4E	GS-220 HQ x2
Palatinose	7.8	6.5	5.9	8.1	4.0	12.1	
Panose	7.1	5.8	5.3	16.9	*	25.6	
L-Phenylalanine	-	-	-	-	31.6	-	
D(+)-Raffinose	7.1	5.8	5.3	16.4	*	20.3	15.1
D(+)-Rhamnose	9.8	8.2	7.4	3.9	4.4	5.5	
D(-)-Ribose	19.4	13.7	9.0	4.8	8.6	5.5	
Rutinose	7.8	6.5	5.8	6.7	*	10.9	
Saccharin Sodium	7.0	5.4	4.3	6.8	*	5.7	
D(-)-Sorbitol	21.6	13.3	7.4	9.8	11.9	7.1	16.6
D(+)-Sorbose	9.7	8.0	7.4	5.1	4.9	7.4	
Stachyose	6.8	5.6	5.0	-	*	36.2	14.3
Stevioside	-	-	-	4.1	*	6.1	
Sucrose	7.5	6.3	5.9	7.9	*	11.9	16.3
$\alpha$ -D-Talose	21.3	12.6	8.8	5.7	8.5	6.5	
Theanderose	2peaks	2peaks	2peaks	2peaks	*	2peaks	
Trehalose	7.6	6.3	5.8	10.9	*	13.3	
Trehalulose	8.9	7.0	6.1	9.5	4.8	11.7	
Xylitol	19.9	13.1	7.9	7.8	10.2	6.1	16.9
Xylobiose	8.2	6.7	6.4	5.7	*	9.1	
D(+)-Xylose	9.2	7.1	7.7	4.6	4.5	6.6	17.5
D-Xylulose	10.6	9.0	8.0	4.1	5.1	5.4	

(-) → cannot be detected (\*) → cannot be separated from solvent peak

Column : Shodex SUGAR SP0810, SC1011, KS-801  
(8.0mmID x 300mm each)  
Eluent : H<sub>2</sub>O  
Flow Rate : 1.0mL/min  
Detector : Shodex RI  
Column Temp. : 80°C

Column : Shodex Asahipak NH2P-50 4E  
(4.6mmID x 250mm)  
Eluent : H<sub>2</sub>O/CH<sub>3</sub>CN=25/75  
Flow Rate : 1.0mL/min  
Detector : Shodex RI  
Column Temp. : 30°C

Column : Shodex SUGAR SZ5532  
(6.0mmID x 150mm)  
Eluent : H<sub>2</sub>O/CH<sub>3</sub>CN=25/75  
Flow Rate : 1.0mL/min  
Detector : Shodex RI  
Column Temp. : 60°C

Column : Shodex Asahipak GS-220 HQ  
(7.5mmID x 300mm) x 2  
Eluent : H<sub>2</sub>O  
Flow Rate : 0.5mL/min  
Detector : Shodex RI  
Column Temp. : 60°C

Column : Shodex SUGAR SC1211  
(6.0mmID x 250mm)  
Eluent : H<sub>2</sub>O/CH<sub>3</sub>CN=65/35  
Flow Rate : 1.0mL/min  
Detector : Shodex RI  
Column Temp. : 70°C

## B. List of Elution Volume of Saccharides and Organic Acids (SH1011)

Sample	Eluent	
	5mM	20mM
Oxalic Acid	5.1	5.6
Stachyose	5.2	5.2
Isomaltotriose	5.3	5.3
Panose	5.4	5.4
Maltotriose	5.4	5.5
Gentiobiose	5.7	5.7
Isomaltose	5.8	5.8
Melibiose	5.9	5.9
Kojibiose	5.9	5.9
Maltose	5.9	5.9
Trehalose	5.9	5.9
Nigerose	5.9	5.9
Palatinit	6.0	6.0
Palatinose	6.0	5.9
Trehalulose	6.0	6.0
Lactose	6.0	6.0
Xylobiose	6.1	6.1
Maltitol	6.1	6.2
D-Glucuronic Acid	6.1	6.2
Citric acid	6.2	6.3
$\alpha$ -Ketoglutaric Acid	6.2	6.9
Maleic Acid	6.2	7.1
Lactitol	6.3	6.3
N-Acetylneuramic Acid	6.3	6.6
Tartaric Acid	6.4	6.6
Glucose	6.8	6.8
Pyruvic Acid	6.9	7.7
Sorbose	6.9	6.9

Sample	Eluent	
	5mM	20mM
myo-Inositol	7.0	7.1
Malic Acid	7.0	7.2
D-Mannose	7.1	7.1
D(+)-Galactose	7.1	7.2
D(+)-Xylose	7.2	7.2
Malonic Acid	7.2	7.5
D(-)-Fructose	7.2	7.2
trans-Aconitic Acid	7.2	7.5
Mannitol	7.4	7.4
D(-)-Sorbitol	7.5	7.5
D(+)-Rhamnose	7.5	7.5
D(+)-Arabinose	7.6	7.7
D(-)-Ribose	7.8	7.8
D-Arabitol	7.8	7.9
Xylitol	8.0	8.0
D(+)-Fucose	8.0	8.0
Succinic Acid	8.3	8.3
meso-Erythritol	8.3	8.3
N-Acetyl- $\alpha$ -D-glucosamine	8.4	8.4
DL-Lactic Acid	8.8	8.9
N-Acetyl- $\alpha$ -D-galactosamine	9.3	9.3
Formic Acid	9.4	9.5
Acetic Acid	10.1	10.2
Adipic Acid	10.4	10.5
Mesaconic Acid	11.0	11.4
L-Pyroglyutamic Acid	11.4	11.7
Propionic Acid	11.7	11.7
Ethanol	14.1	14.1

Column : Shodex SUGAR SH1011 (8.0mmID x 300mm)  
 Eluent : 5mM or 20mM H<sub>2</sub>SO<sub>4</sub> aq.  
 Flow Rate : 0.6mL/min  
 Detector : Shodex RI  
 Column Temp. : 60°C

### C. List of Elution Volume of Organic Acids (KC-811)

Substance	Elution Volume (mL)			
	1mM HClO <sub>4</sub>	2mM HClO <sub>4</sub>	3mM HClO <sub>4</sub>	5mM HClO <sub>4</sub>
Oxalic Acid	9.3	9.4	9.6	9.7
Maleic Acid	9.7	10.2	10.5	11.1
alpha-Ketoglutaric Acid	10.0	10.4	10.9	11.4
Oxalacetic Acid	10.5	11.2	11.8	12.5
Pyruvic Acid	10.5	11.2	11.8	12.5
2-Ketogluconic Acid	10.6	11.1	11.5	11.7
Phosphoric Acid	10.6	11.0	11.3	11.6
Citric Acid	10.8	11.3	11.7	11.8
Tartaric Acid	11.1	11.7	12.1	12.3
Isocitric Acid	11.1	11.5	11.9	11.9
Glucuronic Acid	11.2	11.4	11.7	11.8
trans-Aconitic Acid	11.5	12.4	13.1	13.5
Malonic Acid	11.5	12.4	13.0	13.4
Galacturonic Acid	11.8	12.1	12.4	12.4
Gluconic Acid	12.3	12.5	12.8	12.5
Glyoxylic Acid	12.4	12.9	13.3	13.9
Malic Acid	12.5	13.0	13.4	13.4
alpha-Hydroxyglutaric Acid	13.5	13.9	14.0	14.2
Ascorbic Acid	13.5	13.7	13.9	13.8
Fumaric Acid	14.5	16.1	17.2	17.8
Succinic Acid	15.3	15.5	15.8	15.7
Glycolic Acid	15.6	15.9	16.3	16.2
Itaconic Acid	15.9	16.4	16.8	16.8
Lactic Acid	16.0	16.4	16.7	16.7
2-Hydroxyisobutyric Acid	16.5	16.8	17.1	17.1
Adipic Acid	16.7	19.8	20.2	20.0
beta-Hydroxypropionic Acid	16.9	17.0	17.2	17.1
Formic Acid	17.2	17.6	17.9	17.9
Mesaconic Acid	17.2	19.2	20.5	21.1
Uric Acid	17.9	17.9	18.3	18.0
Pyroglutamic Acid	18.6	20.3	21.0	21.7
Acetic Acid	19.2	19.3	19.4	19.4
Levulinic Acid	20.0	20.2	20.3	20.2
Propionic Acid	22.3	22.4	22.5	22.5
CO <sub>3</sub> <sup>2-</sup> , HCO <sub>3</sub> <sup>3-</sup>	24.7	24.7	24.5	24.7
Isobutyric Acid	25.4	25.5	25.6	25.6
n-Butyric Acid	27.3	27.5	27.6	27.6
Isovaleric Acid	31.9	32.1	32.3	32.2
n-Valeric Acid	38.4	38.6	38.9	38.8

Column : Shodex RSpak KC-811 (8.0mmID x 300mm) x 2  
 Eluent : HClO<sub>4</sub> aq.  
 Flow Rate : 1.0mL/min  
 Column Temp. : 50°C



## D. List of Elution Volume of Organic Acids (KC-811)

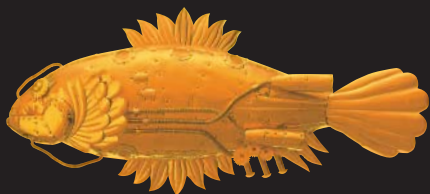
Substance	Elution Volume (mL)					
	30°C	40°C	50°C	60°C	70°C	80°C
Oxalic Acid	9.7	9.6	9.6	9.6	9.5	9.5
Maleic Acid	10.7	10.7	10.5	10.5	10.5	10.4
Phosphoric Acid	11.0	11.1	11.3	11.4	11.5	11.7
$\alpha$ -Ketoglutaric Acid	11.1	11.0	10.9	10.8	10.7	10.6
2-Ketogluconic Acid	11.6	11.5	11.5	11.5	11.4	11.4
Glucuronic Acid	11.7	11.7	11.7	11.7	11.7	*
Pyruvic Acid	11.9	11.8	11.8	11.7	11.7	11.7
Citric Acid	11.9	11.8	11.7	11.6	11.5	11.4
Oxalacetic Acid	12.0	11.9	11.8	11.8	11.7	11.7
Isocitric Acid	12.1	12.0	11.9	11.7	11.6	11.5
Tartaric Acid	12.4	12.2	12.1	12.0	11.9	11.8
Galacturonic Acid	12.5	12.4	12.4	12.3	12.3	12.2
Gluconic Acid	12.8	12.8	12.8	12.8	12.9	12.9
Malonic Acid	13.3	13.1	13.0	12.8	12.7	12.6
trans-Aconitic Acid	14.0	13.5	13.1	12.7	12.5	12.2
Glyoxylic Acid	13.3	13.3	13.3	13.2	13.1	13.1
Malic Acid	13.7	13.5	13.4	13.2	13.0	12.9
Ascorbic Acid	14.1	14.0	13.9	13.7	13.6	*
$\alpha$ -Hydroxyglutaric Acid	14.6	14.3	14.1	13.8	14.0	13.9
Succinic Acid	16.5	16.1	15.8	15.4	15.1	14.8
Glycolic Acid	16.6	16.4	16.3	16.1	15.9	15.7
Lactic Acid	16.8	16.8	16.7	16.5	16.4	16.2
2-Hydroxyisobutyric Acid	17.0	17.1	17.1	16.9	16.8	16.7
$\beta$ -Hydroxypropionic Acid	17.5	17.3	17.2	17.0	16.8	16.6
Itaconic Acid	17.8	17.3	16.8	16.3	15.9	15.5
Formic Acid	18.2	18.0	17.9	17.6	17.5	17.3
Fumaric Acid	19.2	18.0	17.2	16.4	15.8	15.3
Acetic Acid	19.9	19.7	19.4	19.1	18.8	18.6
Uric Acid	20.7	19.3	18.3	17.4	16.8	16.2
Levulinic Acid	22.1	21.1	20.3	19.5	18.9	18.3
Pyroglutamic Acid	22.9	21.8	21.0	20.3	19.8	19.3
Adipic Acid	23.1	21.4	20.2	19.0	18.1	17.3
Propionic Acid	23.5	23.0	22.5	22.0	21.5	20.7
Mesaconic Acid	24.0	21.9	20.5	19.2	18.1	17.3
CO <sub>3</sub> <sup>2-</sup> , HCO <sub>3</sub> <sup>-</sup>	24.3	24.5	24.5	24.5	24.4	24.2
Isobutyric Acid	27.1	26.4	25.6	24.8	24.0	23.1
n-Butyric Acid	29.8	28.7	27.6	26.5	25.5	24.5
Isovaleric Acid	35.7	33.9	32.3	30.6	29.0	27.5
n-Valeric Acid	45.1	41.8	38.9	36.1	33.6	31.3

Column : Shodex RSpak KC-811 (8.0mmID x 300mm) x 2  
 Eluent : 3mM HClO<sub>4</sub> aq.  
 Flow Rate : 1.0mL/min

## Notice

1. Please read the instruction manual accompanying the product in its entirety before using the sugar columns.
2. The specifications for the products are subjected to change without notice for purposes of improvement.
3. No guarantee is offered to figures in this technical paper; those figures should be used just as a reference.
4. Even if no precautions are given in the instruction manual as to the safety or danger of reagents and chemical products, make sure that in handling the products, the usual precautions are taken.
5. The products described herein are not designed for use in clinical examinations in the medical area.

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